

AD_____

Award Number: W81XWH-04-1-0689

TITLE: The Role of SDF-1alpha and CXCR4 in Metastatic Breast Cancer

PRINCIPAL INVESTIGATOR: KM Wahidur Rahman, Ph.D.

CONTRACTING ORGANIZATION: Wayne State University
Detroit, MI 48201-1928

REPORT DATE: August 2006

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE				Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Department of Defense, Washington Headquarters Services, Directorate for Information Operations and Reports (0704-0188), 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302. Respondents should be aware that notwithstanding any other provision of law, no person shall be subject to any penalty for failing to comply with a collection of information if it does not display a currently valid OMB control number. PLEASE DO NOT RETURN YOUR FORM TO THE ABOVE ADDRESS.					
1. REPORT DATE (DD-MM-YYYY) 01-08-2006		2. REPORT TYPE Final		3. DATES COVERED (From - To) 30 JUL 2004 - 29 JUL 2006	
4. TITLE AND SUBTITLE The Role of SDF-1alpha and CXCR4 in Metastatic Breast Cancer				5a. CONTRACT NUMBER	
				5b. GRANT NUMBER W81XWH-04-1-0689	
				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S) KM Wahidur Rahman, Ph.D. E-Mail: kmrahman@med.wayne.edu				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Wayne State University Detroit, MI 48201-1928				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited					
13. SUPPLEMENTARY NOTES					
14. ABSTRACT: A major clinical manifestation of breast cancer patients is the development of bone metastasis. Several lines of experimental evidence suggested that chemokine receptor CXCR4 and its ligand SDF-1α signaling molecules may play an important role in the bone metastasis of breast cancer via activating NF-κB, a key transcription factor. We have previously shown that Indole-3-carbinol (I3C), a natural compound present in vegetables of the genus <i>Brassica</i> can inhibit NF-κB in breast cancer cells. Therefore, we hypothesize that I3C may be able to inhibit the bone metastasis of breast cancer by inhibition of NF-κB, resulting in inhibition of CXCR4/SDF-1α and other NF-κB targeted genes. This is the first time report that we have established an animal model of breast cancer bone metastasis using SCID human (SCID-hu) animal. Using this model, we investigated the effect of I3C on MDA-MB-231 breast cancer cells and experimental MDA-MB-231 bone tumors created by injecting MDA-MB-231 cells into human bone fragments. We found that I3C significantly inhibited MDA-MB-231 bone tumor growth. This correlated well with down regulation of NF-κB. We studied the mRNA expression of CXCR4 and SDF-1α and found that 60μM I3C significantly inhibited SDF-1α and CXCR4 expression in MDA-MB-231 cells. Our hypotheses will be tested using MDA-MB-231 human breast cancer cells growing in the SCID-hu model, wherein we can document the expression profile of specific targeted genes could be altered by I3C treatment. By using a novel siRNA technology to silence the SDF-1α and/or CXCR4 genes, we plan to study how SDF-1α/CXCR4 signaling affects the infiltrating growth of MDA-MB-231 cells to the bone in the SCID-hu model, and to delineate whether the effects are exerted via activation of MMPs, uPA and NF-κB. The data obtained from our experiments will provide novel pathways that could be exploited for the prevention and/or treatment of breast cancer bone metastasis in the clinic. Based on above hypothesis, this proposal seems highly relevant to the mission of the Department of Defense.					
15. SUBJECT TERMS					
16. SECURITY CLASSIFICATION OF:			17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON
a. REPORT	b. ABSTRACT	c. THIS PAGE			USAMRMC
U	U	U	UU	54	19b. TELEPHONE NUMBER (include area code)

Table of Contents

Cover.....	
SF 298.....	2
Introduction.....	4
Body.....	5
Key Research Accomplishments.....	10
Reportable Outcomes.....	10
Conclusions.....	10
References.....	10
Appendices.....	11

Introduction

The majority of patients dying from cancer of the breast have metastasis to the bone or visceral site (1). The present options for treating breast cancer bone metastasis are limited. Therefore, studies to elucidate molecular mechanism(s) of breast cancer bone metastasis and its prevention is of paramount importance. Several lines of experimental evidences have suggested that chemokine receptor CXCR4 and its ligand SDF1- α are over-expressed in metastatic breast cancer cells (2). Moreover, NF- κ B, a key transcription factor may regulate the expression and function of CXCR4 and/or SDF-1 α and activate other metastasis-promoting molecules (uPA, MMPs) in breast cancer cells (3, 4). Thus, agents that directly block the expression of CXCR-4 and/or SDF-1 α signaling partly and due to inhibition of NF- κ B may have great therapeutic potential for treating metastatic breast cancer. It has been shown that Indole-3-carbinol (I3C) can inhibit NF- κ B in breast cancer cells (5-7). Recently, we studied the mRNA expression of CXCR4 and SDF-1 α and found that I3C, significantly inhibited CXCR4/SDF-1 α expression in breast cancer cells. However little is known about how chemopreventive agents inhibits these metastasis-promoting molecules and therefore, we hypothesize that I3C may be able to inhibit the bone metastasis of breast cancer by inhibition of NF- κ B and thereby inhibiting CXCR4/SDF-1 α , and other NF- κ B targeted genes. However, there is no suitable model to investigate the molecular biology of breast cancer metastasis and we trust that SCID-Hu model could be a suitable model to fill this void in our understanding of breast cancer bone metastasis. In this study, we have utilized our recently established the SCID-Hu model of breast cancer bone metastasis to determine the effect of I3C *in vivo*. The purpose of our current investigation: is 1) to determine the effects of I3C on breast cancer cells *in vitro*; 2) to determine the effects of I3C on breast cancer bone tumor growth *in vivo*; 3) to determine the alterations in gene expressions by I3C treatment in both *in vitro* and *in vivo* studies; 4) to compare the gene expression of MDA-MB-231 bone tumors and MDA-MB-231 subcutaneous tumors in SCID mice so as to better understand the molecular mechanism(s) by which I3C exerts its anti-metastatic effect on breast cancer cells; and 5) to determine whether down-regulation of CXCR4 and /or SDF-1 α using siRNA approach will prevent the bone metastasis of MDA-MB-231 cells to the bone in the SCID-Hu model.

Body of Report

The original statement of work in the proposal is listed below:

Task 1: To determine whether metastasis of breast cancer cells to the bone requires both expression of SDF-1 α and CXCR4.

- a. We will determine whether BT-20 and MDA-MB-231 breast cancer cells can establish bone metastatic tumor and show infiltrating growth in the SCID-Hu model but not in the subcutaneous site of the same SCID mouse. We will determine the expression patterns of SDF-1 α and CXCR4 in different cellular compartment (Months 1-3).
- b. We will study whether down-regulation of CXCR4 and/or SDF-1 α using siRNA approach will prevent the bone metastasis of BT-20 and MDA-MB-231 cells to the bone in the SCID-Hu model (Months 1-3).
- c. To explore whether SDF-1 α , CXCR4, MMP-9, MMP-2, IL-8, uPA, and NF- κ B are involved in the differential metastasis and infiltrating growth of BT-20 and MDA-MB-231 cells in the SCID-Hu environment and in the subcutaneous site (Months 4-6).

Task 2: To determine whether I3C can inhibit the bone metastasis of BT-20 and MDA-MB-231 breast cancer cells and whether the mechanism(s) for the possible inhibition is mediated by down-regulation of SDF-1 α , CXCR4, MMPs, IL-8, uPA and NF- κ B.

- a. We will determine whether systematic treatment of the host mice with I3C will prevent the bone metastasis of BT-20 and MDA-MB-231 cells in the SCID-Hu model (Months 7-10).
- b. We will determine whether the possible inhibition of bone metastasis of BT-20 and MDA-MB-231 cells in the SCID-Hu model is associated with down regulation of the expression of CXCR4 in the metastatic tumor cells and SDF-1 α , MMP-2, MMP-9, IL-8, uPA and NF- κ B in the bone marrow stromal cells (Months 10-12).
- c. We will summarize all the data, complete writing of all manuscripts, and submit the final report to DOD (Month 12).

In the last annual report, we have provided the evidences to support the research accomplishments in the last year and indicated that we have completed majority parts of task1 and 2 and plan to complete the rest of task in the extended year. We got an extension for one year with no cost.

We are now reporting the research accomplishments associated with task 1 and 2 outlined in the Statement of Work.

Task 1: To determine whether metastasis of breast cancer cells to the bone requires both expression of SDF-1 α and CXCR4.

A. Development of a novel model of breast cancer grown in the environment of human bone:

This was the critical part of our project to develop an animal model of experimental breast cancer bone metastasis. But there are no other existing animal models in the literature that can be utilized

faithfully for studying the breast cancer bone metastasis using breast cancer cell lines which can grow in the marrow of human bone implanted subcutaneously in to mice. This is the first report that we have established an animal model of experimental breast cancer bone metastasis using SCID-hu animal (**Please see our preliminary data Fig. 1**).

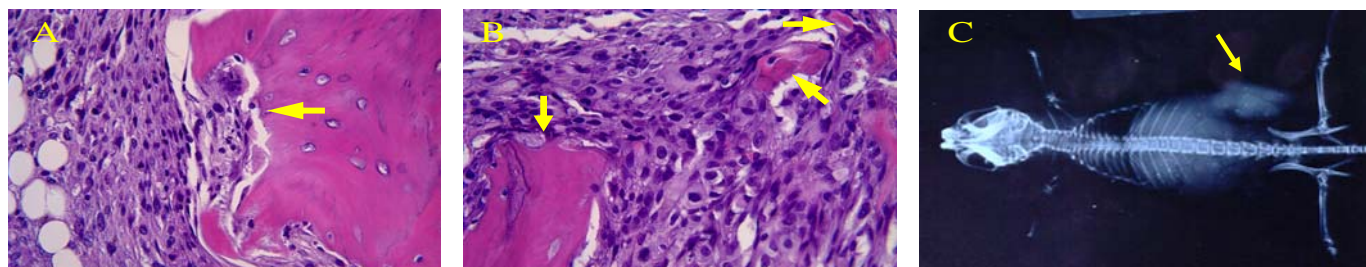


Fig. 1 Typical osteolytic bone metastasis of MDA-MB-231 cells in SCID-Hu model. A: MDA-MB-231 cells which were injected into the implanted human fetal bone showing invasion (arrow); B: Continuous osteolytic and invasive process results in loss of the bone structure. Arrows indicate residuals bone surrounded by invasive tumor cells; C: The osteolytic bone metastasis of MDA-MB-231 cells, which eventually destroy fetal bone structure, as shown by X-ray examination.

Task 2: To determine whether I3C can inhibit the bone metastasis of BT-20 and MDA-MB-231 breast cancer cells and whether the mechanism(s) for the possible inhibition is mediated by down-regulation of SDF-1 α , CXCR4, MMPs, IL-8, uPA and NF- κ B.

A. Effect of I3C during bone cancer metastasis:

Fig. 2 Schematic Design of Animal Experiment

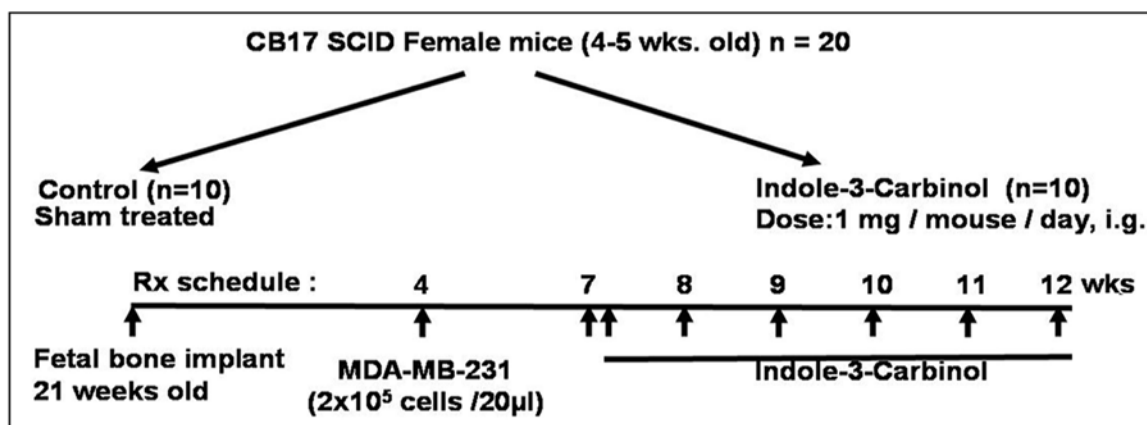


Fig. 2 Flow chart representation of in vivo experimental design and treatment schedule. The human fetal bone was implanted subcutaneously in the right flank of female homozygous CB17 SCID mice through a small skin incision with the open marrow cavity against the mouse muscle. Four weeks after the bone implantation, suspensions of MDA-MB-231 cells (2×10^5 cells) were injected through the mouse skin directly into the marrow of implanted fetal bone. The mice in the intervention group were given I3C (1 mg/day/mouse) by oral gavage everyday for five weeks as soon as the majority of the bone implants began to enlarge (now called a “bone tumor”) as determined by caliper measurements (23rd day after cancer cell injection). The control mice received only saline without I3C. The experiment was terminated 3 months after I3C treatment.

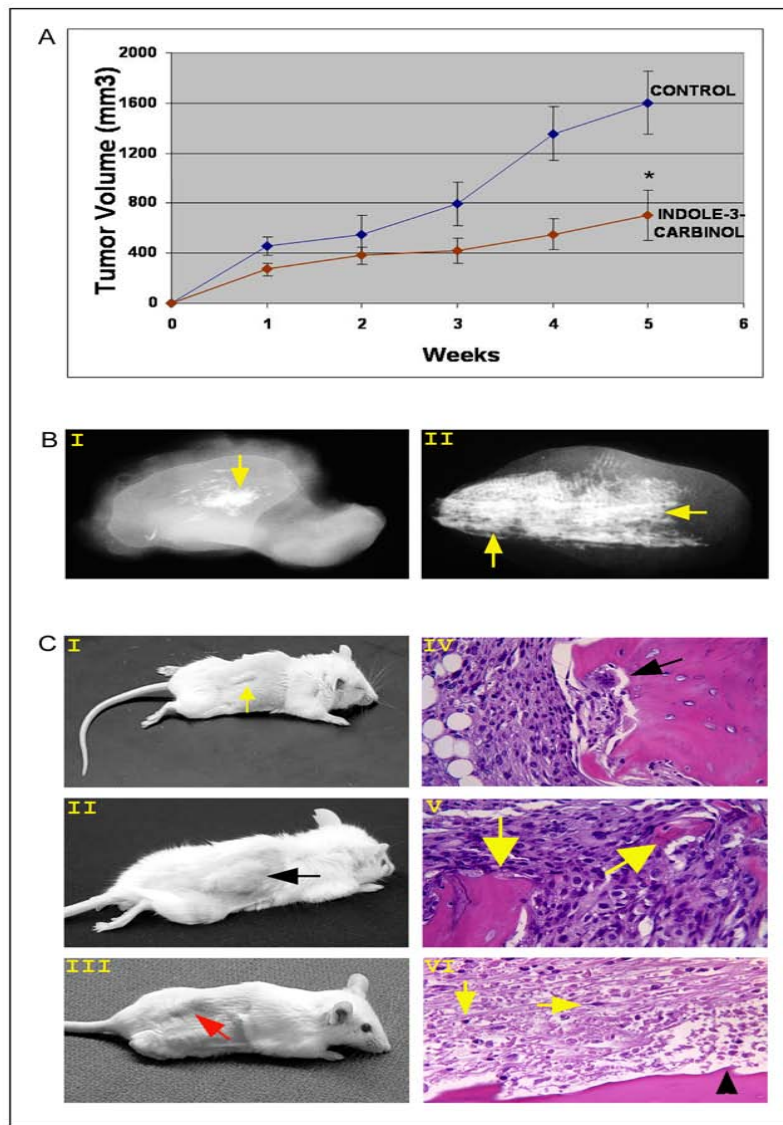


Fig.3 A. Inhibitory effects of I3C on the growth of xenograft bone tumors of MDA-MB-231 cells in SCID-hu mice. A total of 20 mice were divided into 2 groups. Under the experimental conditions, administration of I3C by gavage treatment caused 50% reduction in tumor volume, compared to control group. *, $P < 0.05$. **B.** Ex vivo bone tumor X-ray showing bone osteolysis and tumor growth of MDA-MB-231 cells in the control and I3C-treated. Arrow indicates less residual bone in the control group (B I) relative to I3C-treated (B II). We found that treatment with I3C inhibited tumor and osteolytic bone destruction resulting in more residual bone volume compared to control group (B I). **C.** Typical osteolytic bone metastasis of MDA-MB-231 cells in SCID-hu model. (C I): Human fetal bone subcutaneously implanted in SCID mice; (C II): MDA-MB-231 cells (2×10^5 cells) were injected directly into the marrow of implanted bone. Black arrow represents the site of significant progression of tumor growth in representative untreated control mice; (C III): Representative figure of mice from treatment group photographed at time of sacrifice showing significant regression of tumor growth (red arrow) by I3C treatment; (C IV): MDA-MB-231 cells were injected into the implanted human fetal bone showing invasion (black arrow); (C V): Continuous osteolytic and invasive process resulting in gradual loss of the bone structure. Yellow arrows indicate residual bone surrounded by invasive tumor cells; (C VI): Histological evaluation showing I3C treatment resulted in pronounced death of tumor cells (yellow arrows) and preservation of the bone structure (black arrow head).

Fig 4: I3C abrogates NF- κ B DNA binding activity.

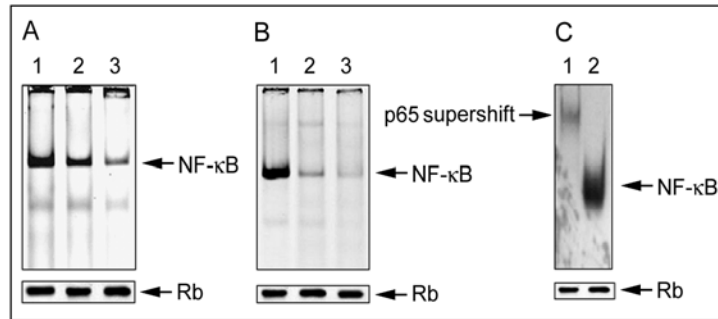


Fig 4 A. I3C abrogates NF- κ B DNA binding activity in MDA-MB-231 breast cancer cells. Cells were treated with 60 or 100 μ M I3C (Lane 2 and 3, respectively) for 72 hours. Nuclear extracts were prepared from control and I3C-treated cells and subjected to analysis for NF- κ B DNA binding activity as measured by electrophoretic mobility shift assay. **B.** MDA-MB-231 cells were grown in the marrow surface of the previously implanted bone in SCID animal. After the termination, tumors were removed and the nuclear protein were subjected to analysis the DNA binding activity of NF- κ B as measured by electrophoretic mobility shift assay (EMSA). Gel shift assay for NF- κ B done on randomly selected frozen tumor tissues obtained from each treatment groups of animals. Group-1 (Lane 1, control animal; Lane 2 and 3, animal received I3C). Results showed that I3C was effective in down-regulating NF- κ B in treated animals relative to control tumors. Arrow indicates NF- κ B DNA binding activity. **C.** The specificity of NF- κ B DNA binding activity was confirmed by supershift assays. Retinoblastoma (Rb) protein level served as nuclear protein loading control.

Fig 5: I3C-induced alterations in metastasis promoting and apoptosis related genes.

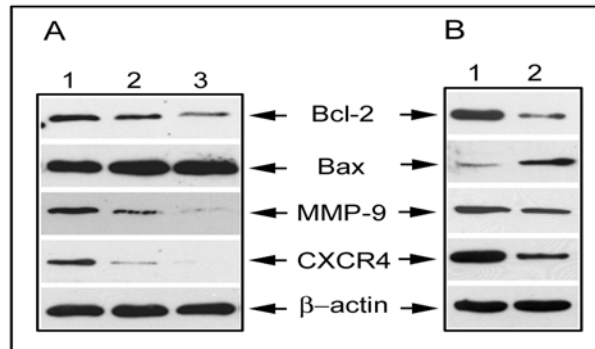


Fig 5 A. I3C down-regulated metastasis related protein such as Bcl-2, Bax, CXCR4 and MMP-9 expression in total cell lysate. Control, MDA-MB-231 cells treated with DMSO; Day 3, cells treated with 60 or 100 μ M I3C (Lane 2 and 3) for 72 hours. Whole cell lysates were prepared and proteins were subjected to Western blot analysis. **B.** Western blot analysis was done on randomly selected frozen tumor tissues obtained from each treatment group of animals. Group-1 (Lane 1, control animal; Lane 2, animal received I3C). Results from Western blot analysis showed that the levels of Bcl-2, Bax, CXCR4 and MMP-9 were significantly down-regulated in I3C treated MDA-MB-231 cells and in animal tumors compared to control group.

Fig 6 . Immunohistochemical staining for CXCR4 and MMP-9

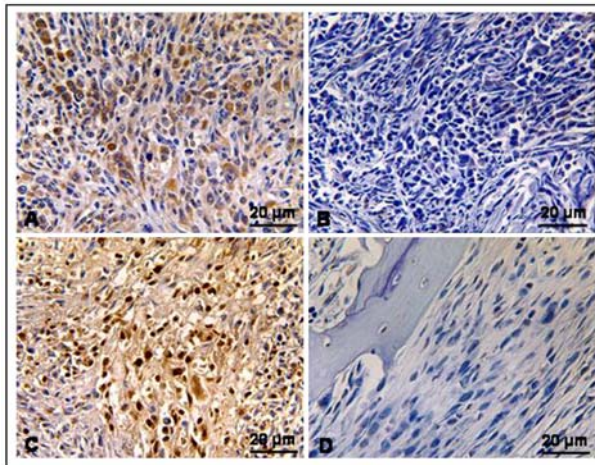


Fig 6. Immunohistochemical staining for CXCR4 and MMP-9 in I3C treated and untreated SCID-Hu model and animal tumors done on randomly selected tumor tissues. Tumor cells in untreated control group show intensive staining of CXCR4 (A) and MMP-9 (C). In the contrast, tumor cells in I3C treated SCID-Hu mice show much weaker staining of CXCR4 (B) and MMP-9 (D). Results from immunohistochemical staining showed that the expression of CXCR4 and MMP-9 was significantly decreased in MDA-MB-231 bone tumors in SCID-hu mice receiving the I3C-containing diet.

Fig 7. I3C inhibits cancer cell invasion and activity of MMP-9

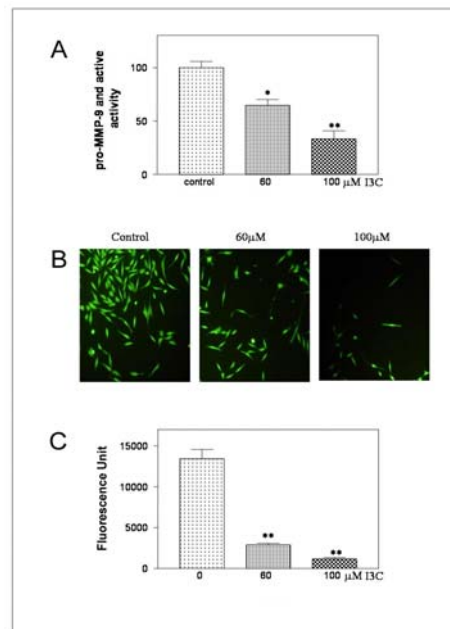


Fig 7 A: MMP-9 expression was down-regulated by I3C. MMP-9 activity assay showed that MMP-9 was down-regulated by I3C in the conditioned medium of MDA-MB-231 breast cancer cells. **B:** I3C induced inhibition of MDA-MB-231 cell invasion. Invasion assay showed that I3C significantly inhibited the invasion of MDA-MB-231 cells through matrigel matrix membrane. **C:** The graphs show the value of fluorescence from the invaded MDA-MB-231 breast cancer cells. The value indicates the comparative amount of invaded MDA-MB-231 breast cancer cells. *, $P < 0.05$; **, $P < 0.01$.

Key Research Accomplishments

- We have established an animal model of experimental human breast cancer bone metastasis.
- Determined the effects of I3C during the growth of MDA-MB-231 bone tumors in an animal model of human breast cancer metastasis.
- Determined the effect of I3C during breast cancer bone metastasis by down-regulating the expression of CXCR-4, MMP-9, NF- κ B and other genes *in vitro* and *in vivo* SCID-hu animals.

Reportable Outcomes

1. KM Wahidur Rahman, Sanjeev Banerjee, Joshua Liao and Fazlul H. Sarkar. The Role of SDF-1 α and CXCR4 in Metastatic Breast Cancer, Proceedings of the Fourth Era of Hope, abstract # 60-21, 2005.
2. Rahman KM, Li Y, Wang Z, Sarkar SH, Sarkar FH (2006). Gene expression profiling revealed survivin as a target of DIM-induced cell growth inhibition and apoptosis in breast cancer cells. Cancer Research, 66(9), 4952-60.
3. KM Wahidur Rahman, Fazlul H. Sarkar, Sanjeev Banerjee, Zhiwei Wang, Dezhong Liao, Xin Hong and Nurul H. Sarkar (2006). Therapeutic intervention of experimental breast cancer bone metastasis by Indole-3-carbinol in SCID-hu mouse model (Communicating to Molecular Cancer Therapeutics, pending minor revision).

Conclusions

- MDA-MB-231 breast cancer cells grew in the human fetal bone and we have established an animal model of experimental breast cancer bone metastasis.
- I3C significantly inhibited the growth of MDA-MB-231 bone tumors in an animal model of human breast cancer metastasis.
- I3C significantly down-regulates the expression of CXCR-4, MMP-9, NF- κ B and other genes *in vitro*.
- I3C inhibited breast cancer bone metastasis by down-regulating the expression of CXCR4, MMP-9 and NF- κ B in SCID-hu animals.
- These results suggest that I3C could be a promising agent for the prevention and/or treatment of breast cancer and its metastasis.

References

1. Coleman RE, Rubens RD. The clinical course of bone metastases from breast cancer. Br J Cancer 1987;55:61-66.
2. Muller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis. Nature 2001;410:50-56.
3. Helbig G, Christopherson KW, 2nd, Bhat-Nakshatri P, et al. NF-kappa B promotes breast cancer cell migration and metastasis by inducing the expression of the chemokine receptor CXCR4. J Biol Chem 2003

4. Nemeth JA, Yousif R, Herzog M, et al. Matrix metalloproteinase activity, bone matrix turnover, and tumor cell proliferation in prostate cancer bone metastasis. *J Natl Cancer Inst* 2002;94:17-25.
5. Rahman KW, Li Y, Sarkar F. Inactivation of Akt and NF- κ B plays important roles during I3c-induced Apoptosis in Breast Cancer cells. *Nutrition and Cancer* 2004;48:84-94.
6. Rahman KW, Li Y, Wang Z, et al. Gene expression profiling revealed survivin as a target of DIM-induced cell growth inhibition and apoptosis in breast cancer cells. *Cancer Research* 2006;In press.
7. Rahman KW, Sarkar FH. Inhibition of nuclear translocation of nuclear factor- κ B contributes to 3,3'-diindolylmethane-induced apoptosis in breast cancer cells. *Cancer Res* 2005;65:364-371.

Appendices

Publications and abstracts during the second year of funding:

1. Rahman KM, Li Y, Wang Z, Sarkar SH, Sarkar FH (2006). Gene expression profiling revealed survivin as a target of DIM-induced cell growth inhibition and apoptosis in breast cancer cells. *Cancer Research*, 66(9), 4952-60.
2. KM Wahidur Rahman, Fazlul H. Sarkar, Sanjeev Banerjee, Zhiwei Wang, Dezhong Liao, Xin Hong and Nurul H. Sarkar (2006). Therapeutic intervention of experimental breast cancer bone metastasis by Indole-3-carbinol in SCID-hu mouse model (Communicating to *Molecular Cancer Therapeutics*, pending minor revision).

Gene Expression Profiling Revealed Survivin as a Target of 3,3'-Diindolylmethane-Induced Cell Growth Inhibition and Apoptosis in Breast Cancer Cells

KM Wahidur Rahman, Yiwei Li, Zhiwei Wang, Sarah H. Sarkar, and Fazlul H. Sarkar

Department of Pathology, Karmanos Cancer Institute, Wayne State University School of Medicine, Detroit, Michigan

Abstract

The phytochemical indole-3-carbinol (I3C), found in cruciferous vegetables, and its major acid-catalyzed reaction product 3,3'-diindolylmethane (DIM) showed anticancer activity mediated by its pleiotropic effects on cell cycle progression, apoptosis, carcinogen bioactivation, and DNA repair. To further elucidate the molecular mechanism(s) by which 3,3'-diindolylmethane exerts its effects on breast cancer cells, we have used microarray gene expression profiling analysis. We found a total of 1,238 genes altered in 3,3'-diindolylmethane-treated cells, among which 550 genes were down-regulated and 688 genes were up-regulated. Clustering analysis showed significant alterations in some genes that are critically involved in the regulation of cell growth, cell cycle, apoptosis, and signal transduction, including down-regulation of survivin. Previous studies have shown that antiapoptotic protein survivin is overexpressed in many human cancers, including breast cancer. However, very little or no information is available regarding the consequence of down-regulation of survivin for cancer therapy. We, therefore, hypothesized that down-regulation of survivin as observed by 3,3'-diindolylmethane could be an important approach for the treatment of breast cancer. We have tested our hypothesis using multiple molecular approaches and found that 3,3'-diindolylmethane inhibited cell growth and induced apoptosis in MDA-MB-231 breast cancer cells by down-regulating survivin, Bcl-2, and cdc25A expression and also caused up-regulation of p21^{WAF1} expression, which could be responsible for cell cycle arrest. Down-regulation of survivin by small interfering RNA before 3,3'-diindolylmethane treatment resulted in enhanced cell growth inhibition and apoptosis, whereas overexpression of survivin by cDNA transfection abrogated 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis. These results suggest that targeting survivin by 3,3'-diindolylmethane could be a new and novel approach for the prevention and/or treatment of breast cancer. (Cancer Res 2006; 66(9): 4952-60)

Introduction

Breast cancer is the second leading cause of cancer-related deaths in women in the United States (1), suggesting that early diagnosis and prevention of this disease is urgently needed. Currently, breast cancer is treated with surgery, chemotherapy, and

radiation therapy or combined modalities with remarkable success. In addition, patients with breast cancer or preneoplastic lesions are also treated with hormonal therapy either for treatment or prevention purposes. Although these treatment modalities are successful, a significant number of patients either do not respond to therapy, or the tumor may recur during therapy and develop metastasis, for which there is limited curative therapy. This inadequate outcome strongly suggests that the evaluation of novel targeted therapeutic agents is urgently needed to improve the treatment outcome of patients diagnosed with this disease.

It has been well known that many genes play important roles in the control of cell growth, differentiation, apoptosis, inflammation, stress response, and many other physiologic processes (2–10). Among those genes, *survivin*, a member of the inhibitor of apoptosis protein (IAP) family, plays important roles in tumorigenesis, progression of breast carcinoma, cell invasion, metastasis, and resistance to chemotherapy (10–15). Several studies have suggested that the antiapoptotic protein survivin is overexpressed in many human cancers, including breast cancer (9, 10). It has also been shown that expression of survivin is associated with cancer cell viability and drug resistance (16). Previous studies in determining the association of survivin with prognosis in breast cancer patients has been controversial (17); thus, the clinical importance of survivin expression remains unclear in patients with breast cancer. We believe that the down-regulation of survivin in breast cancer cells could be a novel therapeutic approach for achieving optimal results in patients with chemoresistant breast cancer. However, very little or no information is currently available regarding the consequence of down-regulation of survivin in cell fate and whether this down-regulation could offer potential therapeutic benefits in breast cancer patients.

Studies from our laboratory and others have shown that 3,3'-diindolylmethane, a major *in vivo* acid-catalyzed condensation product of indole-3-carbinol (I3C), is a potent inhibitor of cell growth and inducer of apoptotic cell death (4, 18–22). However, the comprehensive molecular mechanism(s) by which I3C/3,3'-diindolylmethane inhibits cell growth and apoptosis is still unknown. However, the 3,3'-diindolylmethane's pleiotropic effects on breast cancer cells could be due to alterations in gene expression profiles that are important for cell growth and induction of apoptosis. Thus, understanding the molecular biological properties of 3,3'-diindolylmethane may lead to the clinical development of mechanism-based chemopreventive and/or therapeutic strategies for breast cancer. Microarray gene expression profiling allows examination of the expression of a large number of genes and, in turn, provides an opportunity for determining the effects of anticancer agents on cancer cells. The alterations of gene expression profiles by several anticancer agents have been reported (23, 24). In this study, we used the high-throughput gene chip, which contains 22,215 known genes to better understand the

Requests for reprints: KM Wahidur Rahman, Department of Pathology, Karmanos Cancer Institute, Wayne State University School of Medicine, 9374 Scott Hall, 540 East Canfield, Detroit, MI 48201. Phone: 313-576-8273; Fax: 313-576-8389; E-mail: kmrahman@med.wayne.edu.

©2006 American Association for Cancer Research.
doi:10.1158/0008-5472.CAN-05-3918

precise molecular mechanism(s) by which 3,3'-diindolylmethane exerts its effects on breast cancer cells. We found a total of 1,238 genes altered by 3,3'-diindolylmethane treatment, among which 550 genes were down-regulated and 668 genes were up-regulated, many of which are associated with regulation of cell growth, cell cycle, apoptosis, and intracellular signaling. We also found a significant down-regulation of survivin expression. Based on our results, we further tested and found that 3,3'-diindolylmethane-induced down-regulation of survivin is, in part, mechanistically associated with 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis.

Materials and Methods

Cell culture and growth inhibition. For the present study, we have used human breast epithelial cells MDA-MB-231, which is tumorigenic [American Type Culture Collection (ATCC), Manassas, VA]. It has aggressive invasion capacity and can also grow very well in an animal model (ATCC). MDA-MB-231 was grown in DMEM/F12 (Invitrogen, Carlsbad, CA) supplemented with 10% fetal bovine serum and 1% penicillin and streptomycin in a 5% CO₂ atmosphere at 37°C. 3,3'-Diindolylmethane (LKT Laboratories, St. Paul, MN) was dissolved in DMSO to make 20 mmol/L stock solution and was added directly to the media at different concentrations. Concentration between 30 and 60 µmol/L 3,3'-diindolylmethane seems adequate for this study primarily because results of several studies have indicated that 3,3'-diindolylmethane exhibits promising cancer protective activities, especially against mammary neoplasia (8, 25–28). Moreover, based on these previous studies, including our own study, we have chosen different concentrations of 3,3'-diindolylmethane for this study, which is relevant and achievable *in vivo*. Survivin small interfering RNA (siRNA) and siRNA control were obtained from Santa Cruz Biotechnology (Santa Cruz, CA). LipofectAMINE 2000 was purchased from Invitrogen. Protease inhibitor cocktail, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), and all other chemicals were obtained from Sigma (St. Louis, MO).

Cell growth inhibition studies by MTT assay. For growth inhibition, MDA-MB-231 cells (2×10^3) were seeded in a 96-well culture plate and subsequently treated with 10, 20, 30, 40, and 60 µmol/L 3,3'-diindolylmethane for 24, 48, and 72 hours, whereas control cells received 0.01% DMSO in culture medium. After treatment, the cells were incubated with MTT reagent (0.5 mg/mL; Sigma) at 37°C for 2 hours and then with isopropanol at room temperature for 1 hour. Spectrophotometric absorbance of the samples was determined by an Ultra Multifunctional Microplate Reader (Tecan, Durham, NC). Results were plotted as means \pm SD of three separate experiments having six determinations per experiment for each experimental condition.

Histone/DNA ELISA for detecting apoptosis. The cell apoptosis ELISA detection kit (Roche, Palo Alto, CA) was used to detect apoptosis in breast cancer cells treated with 3,3'-diindolylmethane according to manufacturer's protocol. Briefly, the cytoplasmic histone/DNA fragments from MDA-MB-231 breast cancer cells treated with 40 and 60 µmol/L 3,3'-diindolylmethane or DMSO (vehicle control) for 24, 48, or 72 hours were extracted and bound to immobilized anti-histone antibody. Subsequently, the peroxidase-conjugated anti-DNA antibody was used for the detection of immobilized histone/DNA fragments. After addition of substrate for peroxidase, the spectrophotometric absorbance of the samples was determined by using Ultra Multifunctional Microplate Reader at 405 nm.

Plasmid and transfection. The survivin cDNA plasmid encoding survivin were obtained from Science Reagents (Ipswich, MA). MDA-MB-231 cells were transfected with survivin siRNA and siRNA control, respectively, using LipofectAMINE 2000. MDA-MB-231 cells were transiently transfected with human survivin cDNA. The transfected cells were treated with 40 and 60 µmol/L 3,3'-diindolylmethane for 24, 48, and 72 hours or kept as control. The cell growth and apoptotic cell death of transfected cells with and without treatments were measured using MTT assay and cell apoptosis ELISA Detection kit (Roche), respectively.

cDNA microarray analysis. MDA-MB-231 cells were treated with 40 µmol/L 3,3'-diindolylmethane for 6, 24, and 48 hours. 3,3'-Diindolylmethane is the *in vivo* dimeric product of I3C. The doses of 3,3'-diindolylmethane chosen for the microarray experiment were close to IC₅₀. However, the biological relevance of these doses in relation to prevention or therapy has not been fully evaluated. The rationale for choosing these time points was to capture the expression profiles of early-response genes, genes that may be involved in the onset of growth inhibition and apoptotic processes, and, finally, genes that may be involved during active growth inhibition and apoptosis. Total RNA from each sample was isolated by Trizol (Invitrogen) and purified using the RNeasy Mini kit and RNase-free DNase Set (Qiagen, Valencia, CA) according to the manufacturer's protocols. cDNA for each sample was synthesized using a Superscript cDNA Synthesis kit (Invitrogen) and a T7-(dT)24 primer instead of the oligo-(dT) provided in the kit. Then, the biotin-labeled cRNA was transcribed *in vitro* from cDNA using a BioArray HighYield RNA Transcript Labeling kit (ENZO Biochem, New York, NY) and purified using the RNeasy Mini kit. The purified cRNA was fragmented by incubation in fragmentation buffer [200 mmol/L Tris-acetate (pH 8.1), 500 mmol/L potassium acetate, 150 mmol/L magnesium acetate] at 95°C for 35 minutes and chilled on ice. The fragmented labeled cRNA was applied to the Human Genome U133A Array (Affymetrix, Santa Clara, CA), which contains 22,215 human gene sequence, and hybridized to the probes in the array. After washing and staining, the arrays were scanned using a HP GeneArray Scanner (Hewlett-Packard, Palo Alto, CA). Two independent experiments were done to verify the reproducibility of our results.

Microarray data normalization and analysis. The gene expression levels of samples were normalized and analyzed using Microarray Suite, MicroDB and Data Mining Tool software (Affymetrix). The signal value of the experimental array was multiplied by a normalization factor to make its mean intensity equivalent to the mean intensity of the control array using Microarray Suite software according to manufacturer's protocol. The absolute call (present, marginal, and absent) and average difference of 22,215 gene expressions in a sample and the absolute call difference, fold change, and average difference of gene expressions between two or several samples were identified using the abovementioned software. Statistical analysis of the difference in the mean expression of genes showing a >2-fold change was done repeatedly between treated and untreated samples using *t* tests. Average-linkage hierarchical clustering of the data was applied using the Cluster method (29), and the results were displayed with TreeView (29). The genes showing altered expression were also categorized based on their location and cellular component and reported or suggested biochemical, biological, and molecular functions using Onto-Express (30). Genes that were not annotated or not easily classified were excluded from the functional clustering analysis.

Real-time reverse transcription-PCR analysis for gene expression. To verify the alterations of gene expression at the mRNA level, which appeared on the microarray, we chose 15 representative genes (Table 2) with varying expression profiles for real-time reverse transcription-PCR (RT-PCR) analysis. Two micrograms of total RNA from each sample were subjected to reverse transcription using the Superscript first-strand cDNA synthesis kit (Invitrogen) according to the manufacturer's protocol. Real-time PCR reactions were then carried out in a total of 25 µL of reaction mixture (2 µL of cDNA, 12.5 µL of 2 \times SYBR Green PCR Master Mix, 1.5 µL of each 5 µmol/L forward and reverse primers, and 7.5 µL of H₂O) in an ABI Prism 7700 Sequence Detection System (Applied Biosystems, Foster City, CA). The PCR program was initiated for 10 minutes at 95°C before 40 thermal cycles, each of 15 seconds at 95°C, and 1 minute at 60°C. Data was analyzed according to the comparative C_t method and was normalized by actin expression in each sample. Melting curves for each PCR reaction were generated to ensure the purity of the amplification product.

Western blot analysis. To verify whether the alterations of genes at the level of transcription ultimately result in the alterations at the level of translation, we conducted Western blot analysis for selected genes with varying expression profiles. The MDA-MB-231 cells were treated with 40 and 60 µmol/L 3,3'-diindolylmethane for 6, 12, and 48 hours. After treatment, the cells were lysed, and protein concentration was measured

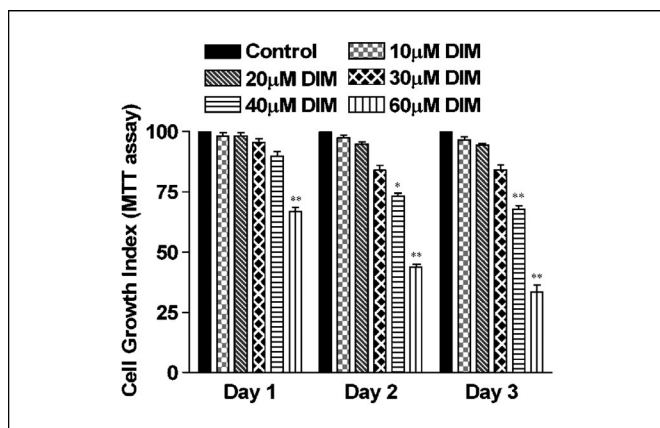


Figure 1. Effects of 3,3'-diindolylmethane (DIM) on the growth of MDA-MB-231 cells. MDA-MB-231 breast cancer cells were treated with 3,3'-diindolylmethane, showing inhibition of cell proliferation in a dose-dependent and time-dependent manner. The proliferation of MDA-MB-231 cells was significantly inhibited by 40 $\mu\text{mol/L}$ 3,3'-diindolylmethane treatment for 48 and 72 hours (30% and 40%, respectively) compared with 60 $\mu\text{mol/L}$ 3,3'-diindolylmethane treatment for 24, 48, and 72 hours (40%, 60%, and 70%, respectively). *, $P < 0.05$; **, $P < 0.01$.

using bicinchoninic acid protein assay (Pierce, Rockford, IL). The proteins were subjected to 10%, 12%, or 14% SDS-PAGE and electrophoretically transferred to nitrocellulose membrane. The membranes were incubated with anti-p21^{WAF1} (1:500; Upstate, Lake Placid, NY), anti-survivin (1:200; R&D Systems, Minneapolis, MN), anti-cell division cycle cdc25A (1:200; Santa Cruz Biotechnology), anti-Bcl-2 (1:250; Calbiochem, San Diego, CA), and anti- β -actin (1:10,000; Sigma) primary antibodies and subsequently incubated with the secondary antibodies conjugated with peroxidase. All

secondary antibodies were obtained from Pierce. The signal was then detected using chemiluminescence detection system (Pierce).

Statistical analysis. The statistical significance was determined using Student's t test, and $P < 0.05$ was considered significant.

Results

Cell growth inhibition by 3,3'-diindolylmethane treatment.

MTT assay showed that the treatment of MDA-MB-231 breast cancer cells with 3,3'-diindolylmethane resulted in a dose- and time-dependent inhibition of cell proliferation (Fig. 1), showing the inhibitory effect of 3,3'-diindolylmethane on MDA-MB-231 cell growth. The proliferation of MDA-MB-231 cells was significantly inhibited by 40 $\mu\text{mol/L}$ 3,3'-diindolylmethane treatment for 48 and 72 hours (30% and 40%, respectively) compared with 60 $\mu\text{mol/L}$ 3,3'-diindolylmethane treatment for 24, 48, and 72 hours (40%, 60%, and 70%, respectively). These results are consistent with our previously published results (8). Inhibition of cell proliferation observed by MTT could be due to altered regulation of several gene expressions by 3,3'-diindolylmethane treatment. Hence, we further investigated the gene expression profile of MDA-MB-231 breast cancer cells treated with 3,3'-diindolylmethane.

Regulation of the mRNA expression by 3,3'-diindolylmethane treatment. The gene expression profile of MDA-MB-231 breast cancer cells treated with 3,3'-diindolylmethane was assessed using cDNA microarray. We found a total of 1,238 genes that showed a >1.5 -fold change after 48 hours of 3,3'-diindolylmethane treatment. Among these genes, 550 genes were down-regulated, and 688 genes were up-regulated in 3,3'-diindolylmethane-treated MDA-MB-231 breast cancer cells. The altered expressions of most

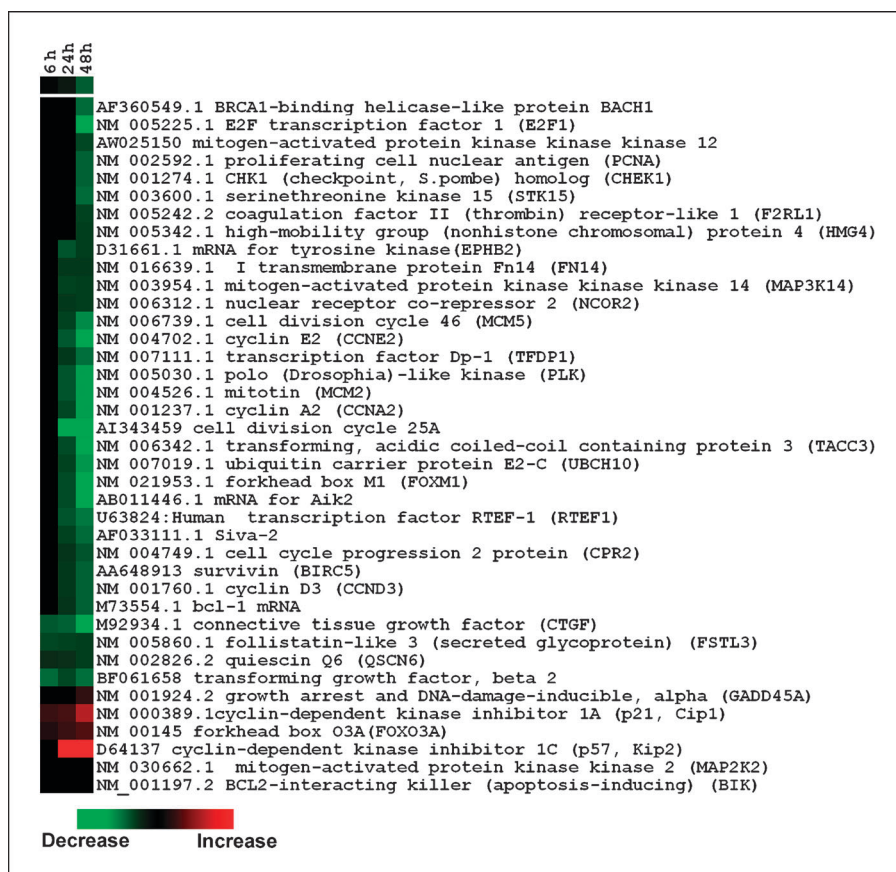


Figure 2. Cluster analysis of genes showing alterations in mRNA expression after 3,3'-diindolylmethane treatment. The alterations of specific and selected genes are shown.

Table 1. Fold changes of specific genes in MDA-MB-231 cells treated with 3,3'-diindolylmethane

Genes	MDA-MB-231		
	6 h	24 h	48 h
Cell cycle, apoptosis, and cell proliferation			
NM_004702.1, cyclin E2 (<i>CCNE2</i>)	NC	-2.2*	-4.2
AI343459, <i>cdc25A</i>	NC	-6.9	-13.9
NM_001760.1, cyclin D3 (<i>CCND3</i>)	NC	-1.5	-2.4
NM_001197.2, BCL2-interacting killer (apoptosis-inducing; BIK)	NC	NC	NC
NM_004749.1, cell cycle progression 2 protein (<i>CPR2</i>)	NC	-1.4	-2.1
NM_006739.1, <i>cdc46</i> (<i>MCM5</i>)	NC	-1.7	-3.4
NM_002592.1, proliferating cell nuclear antigen (<i>PCNA</i>)	NC	NC	-2.5
NM_004526.1, mitotin (<i>MCM2</i>)	NC	-2.1	-4.0
NM_001237.1, cyclin A2 (<i>CCNA2</i>)	NC	-1.8	-4.0
AA648913, survivin (<i>BIRC5</i>)	NC	-1.5	-2.4
AF033111.1, Siva-2	NC	-1.7	-2.6
NM_005860.1, follistatin-like 3 (secreted glycoprotein; <i>FSTL3</i>)	-1.8	-1.7	-1.6
AB011446.1, mRNA for Aik2	NC	-2.0	-4.9
M73554.1, bcl-1 mRNA	NC	-1.4	-2.4
NM_001924.2, growth arrest and DNA damage-inducible, alpha (<i>GADD45A</i>)	NC	NC	1.4
NM_005030.1, polo (<i>Drosophila</i>)-like kinase (<i>PLK</i>)	NC	-2.1	-4.0
D64137, CDK inhibitor 1C (<i>p57^{Kip2}</i>)	NC	22.6 [†]	6.9 [†]
NM_000389.1, CDK inhibitor 1A (<i>p21^{Cip1}</i>)	1.51	1.7 [†]	4.0 [†]
BF061658, transforming growth factor- β 2	-2.6	-1.8	-2.8
NM_00145, <i>FOXO3A</i>	1	1.6	2
Kinase, cell signaling, and cell structure			
NM_003954.1, mitogen-activated protein kinase kinase kinase 14 (<i>MAP3K14</i>)	NC	-1.7	-1.6
NM_030662.1, mitogen-activated protein kinase kinase 2 (<i>MAP2K2</i>)	NC	NC	NC
AW025150, mitogen-activated protein kinase kinase kinase 12	NC	NC	-1.8
D31661.1, mRNA for tyrosine kinase (<i>EPHB2</i>)	NC	-2.1	-1.6
NM_007019.1, ubiquitin carrier protein E2-C (<i>UBCH10</i>)	NC	-1.7	-3.7
Transcription and translation			
NM_021953.1, forkhead box M1 (<i>FOXMI</i>)	NC	-2.0	-4.5
NM_005225.1, E2F transcription factor 1 (<i>E2F1</i>)	NC	NC	-4.2
AF360549.1, BRCA1-binding helicase-like protein <i>BACH1</i>	NC	NC	-2.6
NM_005342.1, high-mobility group (nonhistone chromosomal) protein 4 (<i>HMG4</i>)	1.1	NC	-1.6
U63824, human transcription factor RTEF-1 (<i>RTEF1</i>)	NC	-2.1	-3.0
NM_006312.1, nuclear receptor corepressor 2 (<i>NCOR2</i>)	NC	-1.5	-1.6
NM_007111.1, transcription factor Dp-1 (<i>TFDPI</i>)	NC	-1.5	-2.8
Angiogenesis, metastasis, and invasion			
M92934.1, connective tissue growth factor (<i>CTGF</i>)	-2.2	-2.4	-6.9
NM_006342.1, transforming, acidic coiled-coil containing protein 3 (<i>TACC3</i>)	NC	-2.0	-4.2
NM_016639.1, I transmembrane protein Fn14 (<i>FN14</i>)	NC	-1.6	-1.5
NM_002826.2, quiescin Q6 (<i>QSCN6</i>)	-1.2	-1.3	-1.6
NM_005242.2, coagulation factor II (thrombin) receptor-like 1 (<i>F2RL1</i>)	NC	NC	-1.7
NM_003600.1, serine-threonine kinase 15 (<i>STK15</i>)	NC	NC	-2.6
NM_001274.1, CHK1 (checkpoint, <i>Schizosaccharomyces pombe</i>) homologue (<i>CHEK1</i>)	NC	NC	-2.4

Abbreviation: NC, no change.

*Negative value, decrease.

†Positive value, increase.

genes occurred after only 6 hours of 3,3'-diindolylmethane treatment and were more evident with longer treatment (Fig. 2). After clustering based analysis according to their biological functions, we found down-regulation and up-regulation of several genes, which are related to apoptosis, angiogenesis, tumor cell invasion, and metastasis (Fig. 2; Table 1).

Target confirmation by real-time RT-PCR. To confirm these alterations in gene expression, we conducted RT-PCR analysis of selected genes (Table 2). The results of RT-PCR analysis for these

selected genes (*BACH1*, *BIRC5*, *CCNE2*, *cdc25A*, *F2F1*, *FOXMI*, *MCM5*, *p21^{Cip1}*, *p57^{Kip2}*, *PCNA*, *TACC3*, and *UBCH10*) were in direct agreement with the microarray data (Fig. 3A; Table 1). The same alternations at the mRNA levels were observed by RT-PCR analysis after 3,3'-diindolylmethane treatment, although the fold change in the expression level was not exactly similar between these two different analytic methods (Fig. 3A; Table 1). To further verify the regulation of gene expression at the protein level by 3,3'-diindolylmethane treatment, we narrowed down few genes from the

RT-PCR analysis and conducted Western blot analysis only for selected few genes, which are the main focus of this study and for those genes that are critically important for the inhibition of cell growth and induction of apoptosis. Western blot analysis showed that the protein levels of BIRC5 (survivin), Bcl-2, and cdc25A were down-regulated, and p21^{WAF1} was up-regulated after 3,3'-diindolylmethane treatment (Fig. 3B). These results suggest that 3,3'-diindolylmethane regulated the transcription of the genes involved in apoptosis and cell cycle arrest, angiogenesis, tumor cell invasion, and metastasis, and these results are consistent with our previously published data in prostate cancer cells (3).

Regulation of genes involved in cell cycle and apoptotic process. After clustering based on biological function using OntoExpress and GenMAPP computerized analysis, we found that in MDA-MB-231 breast cancer cells, 3,3'-diindolylmethane down-regulated the expression of some genes that are critically involved in the regulation of cell proliferation and cell cycle (Table 1; Figs. 2 and 4). In contrast, 3,3'-diindolylmethane up-regulated the expression of some genes that are related to induction of apoptosis and cell cycle arrest (Table 1; Figs. 2 and 4). Western blot analysis showed that the protein levels of p21^{WAF1} in MDA-MB-231 cells treated for 48 hours with 3,3'-diindolylmethane were up-regulated, whereas survivin, Bcl-2, and cdc25A were down-regulated (Fig. 3B). These results are novel and have not been previously shown. These results are also in direct agreement with the microarray and RT-PCR data. In addition to the effects of 3,3'-diindolylmethane on cell cycle and apoptosis, 3,3'-diindolylmethane also showed the

regulation of genes related to signal transduction, transcription factor, oncogenesis, and tumor suppression (Table 1; Fig. 2). However, among these genes, *survivin* may have different roles in apoptosis in breast cancer, and because the clinical importance of survivin expression remains unclear in patients with breast cancer, we further investigated the mechanistic role of survivin during 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis.

Down-regulation of survivin expression by siRNA promotes 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis. We used Western blot analysis to detect the protein level of survivin. Intracellular survivin was down-regulated in survivin siRNA-transfected MDA-MB-231 cells compared with siRNA control-transfected cells (Fig. 5). Down-regulation of survivin expression significantly attenuated cell growth inhibition induced by 3,3'-diindolylmethane (Fig. 6A). We found that treatment of cells with 3,3'-diindolylmethane or survivin siRNA alone for 72 hours generally caused 60% to 70% of growth inhibition in MDA-MB-231 cells compared with control. However, 3,3'-diindolylmethane plus survivin siRNA resulted in ~90% growth inhibition compared with control. Survivin siRNA-transfected MDA-MB-231 cells were significantly more sensitive to spontaneous and 3,3'-diindolylmethane-induced apoptosis (Fig. 6B). These results suggest that 3,3'-diindolylmethane plus survivin siRNA promotes cell growth inhibition and apoptosis to a greater degree compared with either agent alone.

Overexpression of survivin by cDNA transfection reduced 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis. Overexpression of survivin by cDNA transfection rescued 3,3'-diindolylmethane-induced cell growth inhibition and abrogated 3,3'-diindolylmethane-induced apoptosis to a certain degree (Fig. 6C and D). We found that treatment of cells with survivin cDNA for 72 hours promotes ~90% of cell growth in MDA-MB-231 cells compared with control. However, 3,3'-diindolylmethane plus survivin cDNA resulted in ~35% growth inhibition compared with control (Fig. 6C). These results provide evidence for a potential role of survivin during 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis in MDA-MB-231 cells.

Discussion

In the present study, we showed that 3,3'-diindolylmethane elicits a significant effect on growth inhibition and induction of apoptotic processes in MDA-MB-231 breast cancer cells mediated by alterations in the gene expression of cell cycle and apoptosis regulatory genes as shown by microarray analysis. Among these genes, we found down-regulation of several genes, such as *BACH1*, *BIRC5* (survivin), *CCNE2*, *CDC25A*, *Bcl-2*, *E2F1*, *FOXM1*, *MCM5*, *PCNA*, *TACC3*, *UBCH10*, *Aik2*, and *CTGF*, and up-regulation of several genes, such as *p21^{Cip1}*, *p57^{Kip2}*, and *FOXO3A*, in 3,3'-diindolylmethane-treated MDA-MB-231 cells that are related to the inhibition of cell growth and induction of apoptosis. To further elucidate the mechanisms of increased apoptosis induced by 3,3'-diindolylmethane, we found that survivin is playing an important role during 3,3'-diindolylmethane-induced cell death.

Our results also showed that cyclins (cyclin A2 and cyclin E2) and CDCs (cdc25A and cdc45) were down-regulated in 3,3'-diindolylmethane-treated breast cancer cells, whereas CDK inhibitors (p21^{WAF1} and p57) were up-regulated, suggesting that 3,3'-diindolylmethane inhibited the growth of breast cancer cell through the arrest of cell cycle and inhibition of proliferation (Fig. 4). Similar effects on the cell cycle were also observed in

Table 2. The primer used for real-time RT-PCR analysis

Genes	Primer sequence
<i>CCNE2</i>	GAATGTCAAGACGAAGTA ATGAACATATCTGCTCTC
<i>cdc25A</i>	ACACAGCAACTGCCATCTCCAG GCCAGCCTCCTTACCATCACG
<i>MCM5</i>	GACCATCTCTATCGCCAAG CTCCTCATTGTGCTCATCC
<i>BIRC5</i>	GCTTTCAGGTGCTGGTAG GATGTGGATCTCGGCTTC
<i>PCNA</i>	CCTGTAGCGGCGTGTGTG CGTTGATGAGGTCCCTTGAGTG
<i>Aik2</i>	ACTTCGGCTGGTCTGTCC ATAGGTCTCGTTGTGTGATGC
<i>p57^{Kip2}</i>	GGACGAGACAGGCGAACC AGAGGACAGCGAGAAGAAGG
<i>p21^{Cip1}</i>	TCCAGCGACCTTCCTCATCCAC TCCATAGCCTCTACTGCCACCATC
<i>UBCH10</i>	TAGGAGAACCCAAACATTGATA AGACGACACAAGGACAGG
<i>FOXMI</i>	AACCGCTACTTGACATTGG GCAGTGGCTTCATCTTCC
<i>E2F1</i>	CAAGAAGTCCAAGAACCACATCC CTGCTGCTCGCTCTCCTG
<i>BACH1</i>	TTTGGGACACGCACACAC ACCGACTACCTCAGGATGG
<i>CTGF</i>	ACCAATGACAACGCCCTCTG TTGCCCTTCTTAATGTTCTCTTCC
<i>TACC3</i>	ATCGTCTGTTCTTCGTGTG AGTCCAAGGGTGTCTATCC
<i>β-Actin</i>	CCACACTGTGCCCATCTACG AGGATCTTCATGAGGTAGTCAGTCAG

3,3'-diindolylmethane-treated prostate cancer cells (3). Several studies have shown that CDCs control the molecules related to the cell cycle, and they are vital and important for the initiation and progression of the consecutive phase of the cell cycle (31, 32). It has been found that cyclins binds to cyclin-dependent protein kinase (CDK) and CDCs to control the cell cycle process (33–36). The CDK inhibitors, including p21^{WAF1}, p27, and p57, have been shown to arrest the cell cycle and inhibit the growth of cancer cells (37–39). Inhibition of cell growth by 3,3'-diindolylmethane could also be due to the induction of apoptosis in addition to cell cycle arrest. Moreover, after analyzing microarray data, we focused on determining whether 3,3'-diindolylmethane could induce apoptosis by inhibiting the expression of survivin, which may be critically important for cell survival and cell death.

Survivin is an IAP, which is overexpressed in human cancer cells and critically needed for regulation of the balance among cell proliferation, differentiation, and apoptosis (17). It is known that nuclear factor- κ B (NF- κ B), a key transcription factor, up-regulates the expression and function of survivin in breast cancer cells (40). It has also been shown that expression of antiapoptotic protein

survivin is associated with cancer cell viability and drug resistance (10). Previous studies in determining the association of survivin with prognosis in breast cancer patients has been controversial (17); thus, the clinical importance of survivin expression remains unclear in patients with breast cancer. However, down-regulation of survivin signaling may be a novel approach in breast cancer therapy. Because of the critical role of survivin in cell proliferation and apoptosis, we explored whether survivin could be a molecular target for 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis. In this study, we showed, for the first time that 3,3'-diindolylmethane down-regulates survivin expression along with other antiapoptotic molecules. We also found that down-regulation of survivin by siRNA together with 3,3'-diindolylmethane treatment caused cell growth inhibition and apoptosis to a greater degree in breast cancer cells compared with either agent alone. Over-expression of survivin by survivin cDNA transfection abrogated 3,3'-diindolylmethane-induced apoptosis to certain degree. Therefore, we strongly believe that down-regulation of survivin is mechanistically linked with 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis.

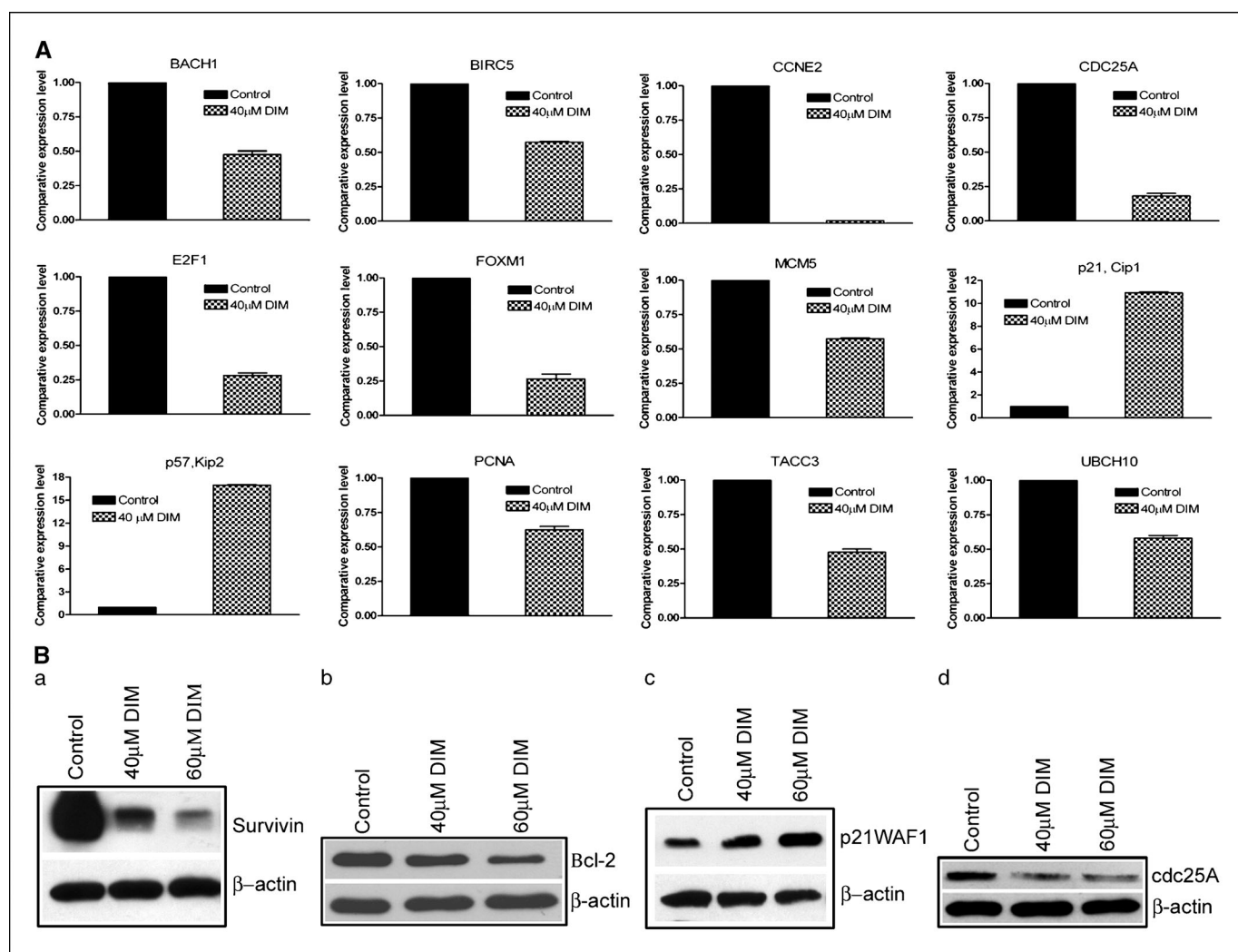


Figure 3. A, real-time RT-PCR amplification value showing the altered expression of specific genes from RNA of 3,3'-diindolylmethane (DIM)-treated MDA-MB-231 cells. B, Western blot analysis of selected gene expression in 3,3'-diindolylmethane-treated MDA-MB-231 cells at the protein level. a, expression of survivin was down-regulated with 3,3'-diindolylmethane treatment. b, expression of Bcl-2 was down-regulated with 3,3'-diindolylmethane treatment. c, expression of p21^{WAF1} was up-regulated with 3,3'-diindolylmethane treatment. d, expression of cdc25A was down-regulated with 3,3'-diindolylmethane treatment.

Author: Modified by Rahman
 Maintained by: WSU
 E-mail: kmrahman@med.wayne.edu
 Last modified: 9/27/2005
 Right-click here to see notes

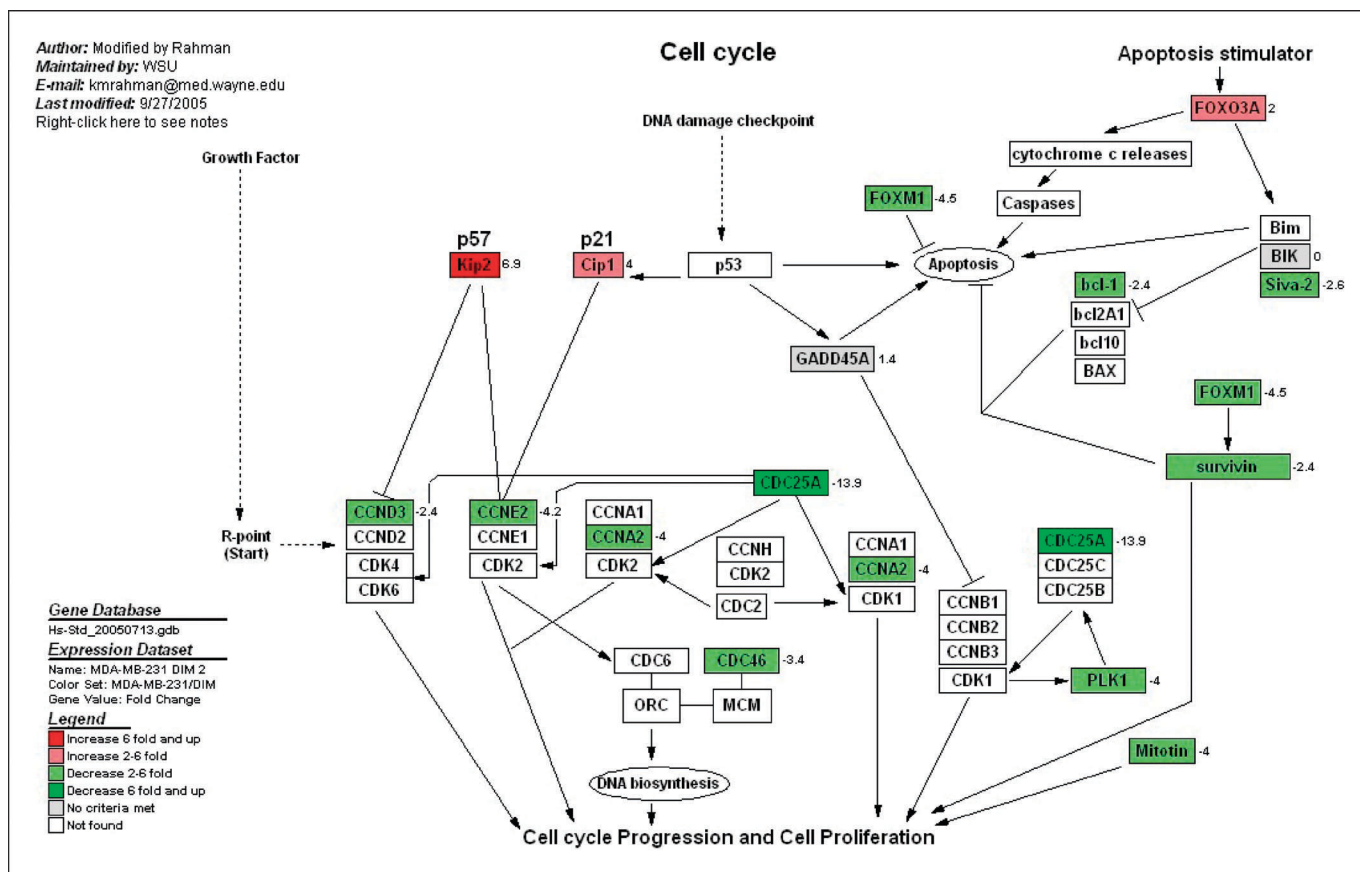


Figure 4. Effect of 3,3'-diindolylmethane on cell cycle and apoptosis pathway-related gene expression as analyzed and visualized by GenMAP software integrated with cDNA microarray data (positive value, increase in fold change; negative value, decrease in fold change).

It has been reported that ectopic expression of survivin, an inhibitor of apoptosis, confers resistance to apoptosis to a variety of stimuli, and survivin is one of the most abundantly overexpressed genes in human tumors (41, 42). Genetic and biochemical data indicate that survivin functions in a unique cell division checkpoint that ensures the apoptotic demise of genetically unstable cells (41). Based on these findings, survivin has been proposed as a suitable target for drugs that can restore the apoptotic program in human tumors. A recent study has indicated that anti-survivin oligonucleotides induced apoptosis in mesothelinoma cells, suggesting that down-regulation of survivin seems to be an effective therapy for mesothelinoma (42). Another study has shown that survivin is expressed in the G₂-M phase of the cell cycle in a cycle-regulated manner (41). It has been shown that GADD45A promotes apoptosis and regulates G₂-M arrest (43), and in our study, we found that GADD45A and FOXO3A expression was up-regulated by 3,3'-diindolylmethane treatment, which could be associated with apoptotic mechanisms (44, 45). Induction of apoptosis by FOXO3A has been correlated with the disruption of mitochondrial member integrity, cytochrome *c* release, and up-regulation of BIM (46). The induction of apoptosis mediated by these molecules, including survivin, could be important molecular mechanism(s) by which 3,3'-diindolylmethane exerts its growth-inhibitory effects on breast cancer cells.

We also found that 3,3'-diindolylmethane down-regulated the expression of the forkhead transcription factor FOXM1, which correlates with proliferative status in a variety of normal and

transformed cell types (47). Elevated expression of FOXM1 has been noted in both hepatocellular carcinoma and basal cell carcinoma (47). It has been indicated that FOXM1 is significantly elevated in primary breast cancer (47). Our microarray analysis showed that FOXM1 regulates genes, including *survivin*, that are essential for mitosis. Loss of FOXM1 expression generates mitotic spindle defects, delays cells in mitosis, and induces mitotic catastrophe (47). At the beginning of mitosis, survivin associates

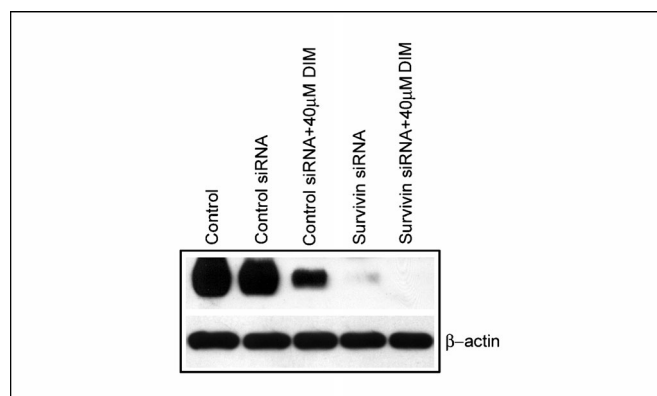
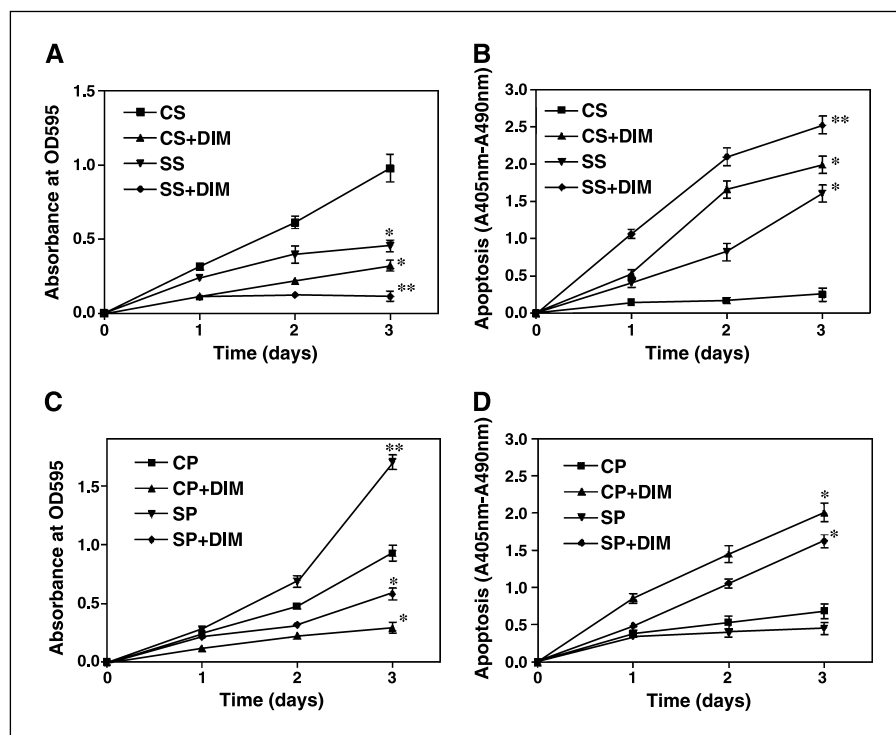


Figure 5. MDA-MB-231 breast cancer cell growth inhibition and cell death induced by survivin siRNA and 3,3'-diindolylmethane. Survivin expression was down-regulated by 3,3'-diindolylmethane and survivin siRNA, resulting in cell death.

Figure 6. MDA-MB-231 breast cancer cell growth inhibition and cell death by survivin cDNA, survivin siRNA, and 3,3'-diindolylmethane (DIM). CS, control siRNA; SS, survivin siRNA; CP, control plasmid; SP, survivin plasmid. **A**, down-regulation of survivin expression significantly inhibited cell growth. 3,3'-Diindolylmethane plus survivin siRNA inhibited cell growth to a greater degree compared with 3,3'-diindolylmethane or survivin alone. **B**, MDA-MB-231 breast cancer cell death induced by survivin siRNA and 3,3'-diindolylmethane. Down-regulation of survivin expression significantly increased apoptosis induced by 3,3'-diindolylmethane. Survivin siRNA-transfected MDA-MB-231 cells were significantly more sensitive to spontaneous and 3,3'-diindolylmethane-induced apoptosis. **C**, overexpression of survivin expression significantly promoted cell growth. Overexpression in survivin rescued breast cancer cells from 3,3'-diindolylmethane-induced cell growth inhibition. **D**, overexpression of survivin by survivin cDNA transfection abrogated 3,3'-diindolylmethane-induced apoptosis. *, $P < 0.05$; **, $P < 0.01$.



with microtubules of the mitotic spindle on a specific and saturable reaction that is regulated by microtubule dynamics (48). Our present findings indicated that inhibition of FOXM1 expression by 3,3'-diindolylmethane, which may disrupt survivin-microtubule interactions, resulted in the loss of survivin's antiapoptosis function and increased caspase-3 activity, a mechanism involved in cell death during mitosis. In contrast, 3,3'-diindolylmethane up-regulated the expression of tumor-suppressor genes, including *p21^{WAF1}* and *p57*, which may inhibit survivin expression. These results are novel and suggest that 3,3'-diindolylmethane may induce the apoptotic pathway by inhibiting cancer cell growth and survival through inhibition of transcription and oncogenesis (hypothetical diagram as in Fig. 7).

Several other potential mechanisms could explain the regulation of survivin associated with inhibition of cell proliferation and

induction of apoptosis. It is known that NF- κ B, a transcription factor, plays important roles in the control of cell growth, differentiation, and apoptosis (11, 12, 14, 16, 49, 50). Several studies have suggested that the activation of NF- κ B up-regulates the expression of its downstream genes, such as *Bcl-2/Bcl-X_L* and *survivin*, which are involved in cancer cell invasion, metastasis, and resistance to chemotherapy (16, 51). We have previously found that 3,3'-diindolylmethane is a potent inhibitor of NF- κ B DNA-binding activity in breast cancer cells (8). Thus, 3,3'-diindolylmethane induced down-regulation of survivin expression could be partly due to inhibition of NF- κ B activity (hypothetical diagram as in Fig. 7). We, therefore, believe that 3,3'-diindolylmethane could inhibit NF- κ B expression, resulting in the down-regulation of its target genes, including *survivin*, and causing cell growth inhibition and apoptosis.

Taken together, we believe that targeting survivin by 3,3'-diindolylmethane could be a novel approach for the prevention and/or treatment of breast cancer. Moreover, the down-regulation of survivin by 3,3'-diindolylmethane could also be a useful strategy for chemosensitization of metastatic breast cancer cells to standard therapies. However, further in-depth investigations are needed to establish the cause and effect relationship of *survivin* gene regulation and 3,3'-diindolylmethane-induced cell growth inhibition and apoptosis in breast cancer in animal and human models.

Acknowledgments

Received 11/1/2005; revised 1/5/2006; accepted 2/16/2006.

Grant support: Two concept awards funded by Department of Defense grants W81XWH-04-1-0689 and W81XWH-05-1-0505 (KM Wahidur Rahman).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

We thank Carrie Koerner for editorial assistance.

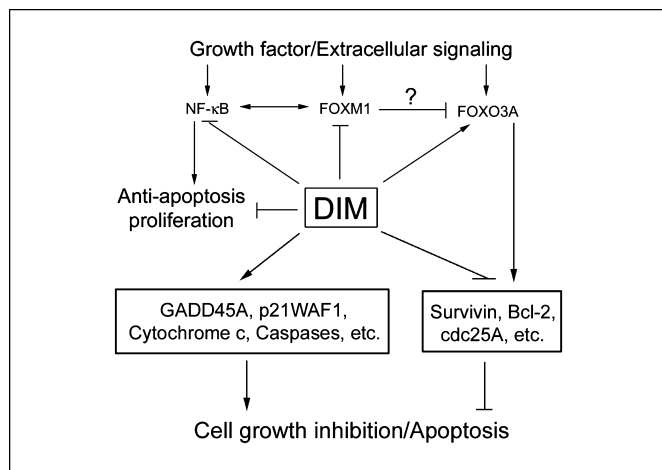


Figure 7. A schematic representation showing 3,3'-diindolylmethane (DIM)-induced cell growth inhibition and apoptosis in breast cancer cells.

References

- Seymour JD, Calle EE, Flagg EW, Coates RJ, Ford ES, Thun MJ. Diet Quality Index as a predictor of short-term mortality in the American Cancer Society Cancer Prevention Study II Nutrition Cohort. *Am J Epidemiol* 2003;157:980–8.
- Li Y, Che M, Bhagat S, et al. Regulation of gene expression and inhibition of experimental prostate cancer bone metastasis by dietary genistein. *Neoplasia* 2004;6:354–63.
- Li Y, Li X, Sarkar FH. Gene Expression profiles of I3C- and DIM-treated PC3 human prostate cancer cells determined by cDNA microarray analysis. *J Nutr* 2003;133:1011–9.
- Rahman KM, Li Y, Sarkar F. Inactivation of Akt and NF- κ B play important roles during I3C-induced apoptosis in breast cancer cells. *Nutr Cancer* 2004;48:84–94.
- Rahman KM, Aranha O, Glazyrin A, Chinni SR, Sarkar FH. Translocation of Bax to mitochondria induces apoptotic cell death in indole-3-carbinol (I3C) treated breast cancer cells. *Oncogene* 2000;19:5764–71.
- Rahman KM, Sarkar FH. Steroid hormone mimics: molecular mechanisms of cell growth and apoptosis in normal and malignant mammary epithelial cells. *J Steroid Biochem Mol Biol* 2002;80:191–201.
- Rahman KM, Aranha O, Sarkar FH. Indole-3-carbinol (I3C) induces apoptosis in tumorigenic but not in non-tumorigenic breast epithelial cells. *Nutr Cancer* 2003;45:101–12.
- Rahman KM, Sarkar FH. Inhibition of nuclear translocation of nuclear factor- κ B contributes to 3,3'-diindolylmethane-induced apoptosis in breast cancer cells. *Cancer Res* 2005;65:364–71.
- Tsuji N, Furuse K, Asanuma K, et al. Mutations of the p53 gene and loss of heterozygosity at chromosome 17p13.1 are associated with increased survivin expression in breast cancer. *Breast Cancer Res Treat* 2004;87:23–31.
- Wu J, Ling X, Pan D, et al. Molecular mechanism of inhibition of survivin transcription by the GC-rich sequence selective DNA-binding antitumor agent, hedamycin: evidence of survivin downregulation associated with drug sensitivity. *J Biol Chem* 2005;280:9745–51.
- Biswas DK, Dai SC, Cruz A, Weiser B, Graner E, Pardee AB. The nuclear factor κ B (NF- κ B): a potential therapeutic target for estrogen receptor negative breast cancers. *Proc Natl Acad Sci U S A* 2001;98:10386–91.
- Hideshima T, Chauhan D, Richardson P, et al. NF- κ B as a therapeutic target in multiple myeloma. *J Biol Chem* 2002;277:16639–47.
- Ikeguchi M, Kaibara N. Changes in survivin messenger RNA level during cisplatin treatment in gastric cancer. *Int J Mol Med* 2001;8:661–6.
- Orlowski RZ, Baldwin AS, Jr. NF- κ B as a therapeutic target in cancer. *Trends Mol Med* 2002;8:385–9.
- Zhang SQ, Qiang SY, Yang WB, Jiang JT, Ji ZZ. Expression of survivin in different stages of carcinogenesis and progression of breast cancer. *Ai Zheng* 2004;23:697–700.
- Haefliger B. NF- κ B: arresting a major culprit in cancer. *Drug Discov Today* 2002;7:653–63.
- Span PN, Sweep FC, Wiegand ET, et al. Survivin is an independent prognostic marker for risk stratification of breast cancer patients. *Clin Chem* 2004;50:1986–93.
- Ashok BT, Chen YG, Liu X, et al. Multiple molecular targets of indole-3-carbinol, a chemopreventive anti-estrogen in breast cancer. *Eur J Cancer Prev* 2002;11 Suppl 2:S86–93.
- Chang X, Tou JC, Hong C, et al. 3,3'-Diindolylmethane inhibits angiogenesis and the growth of transplantable human breast carcinoma in athymic mice. *Carcinogenesis* 2005;26:771–8.
- Hong C, Firestone GL, Bjeldanes LF. Bcl-2 family-mediated apoptotic effects of 3,3'-diindolylmethane (DIM) in human breast cancer cells. *Biochem Pharmacol* 2002;63:1085–97.
- Hong C, Kim HA, Firestone GL, Bjeldanes LF. 3,3'-Diindolylmethane (DIM) induces a G(1) cell cycle arrest in human breast cancer cells that is accompanied by Sp1-mediated activation of p21(WAF1/CIP1) expression. *Carcinogenesis* 2002;23:1297–305.
- Howells LM, Gallacher-Horley B, Houghton CE, Manson MM, Hudson EA. Indole-3-carbinol inhibits protein kinase B/Akt and induces apoptosis in the human breast tumor cell line MDA MB468 but not in the nontumorigenic HBL100 line. *Mol Cancer Ther* 2002;1:1161–72.
- Kudoh K, Ramanna M, Ravatn R, et al. Monitoring the expression profiles of doxorubicin-induced and doxorubicin-resistant cancer cells by cDNA microarray. *Cancer Res* 2000;60:4161–6.
- Zembutsu H, Ohnishi Y, Tsunoda T, et al. Genome-wide cDNA microarray screening to correlate gene expression profiles with sensitivity of 85 human cancer xenografts to anticancer drugs. *Cancer Res* 2002;62:518–27.
- Anderton MJ, Jukes R, Lamb JH, et al. Liquid chromatographic assay for the simultaneous determination of indole-3-carbinol and its acid condensation products in plasma. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003;787:281–91.
- Shilling AD, Carlson DB, Katchamart S, Williams DE. 3,3'-diindolylmethane, a major condensation product of indole-3-carbinol, is a potent estrogen in the rainbow trout. *Toxicol Appl Pharmacol* 2001;170:191–200.
- Sizemore N, Leung S, Stark GR. Activation of phosphatidylinositol 3-kinase in response to interleukin-1 leads to phosphorylation and activation of the NF- κ B p65/RelA subunit. *Mol Cell Biol* 1999;19:4798–805.
- Stresser DM, Blanchard AP, Turner SD, et al. Substrate-dependent modulation of CYP3A4 catalytic activity: analysis of 27 test compounds with four fluorometric substrates. *Drug Metab Dispos* 2000;28:1440–8.
- Eisen MB, Spellman PT, Brown PO, Botstein D. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci U S A* 1998;95:14863–8.
- Khatri P, Draghici S, Ostermeier GC, Krawetz SA. Profiling gene expression using onto-express. *Genomics* 2002;79:266–70.
- Nilsson I, Hoffmann I. Cell cycle regulation by the Cdc25 phosphatase family. *Prog Cell Cycle Res* 2000;4:107–14.
- Poggioli GJ, Dermody TS, Tyler KL. Reovirus-induced signals-dependent G(2)/M phase cell cycle arrest is associated with inhibition of p34(cdc2). *J Virol* 2001;75:7429–34.
- Brechot C. Oncogenic activation of cyclin A. *Curr Opin Genet Dev* 1993;3:11–8.
- Erlanson M, Landberg G. Prognostic implications of p27 and cyclin E protein contents in malignant lymphomas. *Leuk Lymphoma* 2001;40:461–70.
- Obaya AJ, Sedivy JM. Regulation of cyclin-Cdk activity in mammalian cells. *Cell Mol Life Sci* 2002;59:126–42.
- Sherr CJ. D-type cyclins. *Trends Biochem Sci* 1995;20:187–90.
- el-Deiry WS, Tokino T, Waldman T, et al. Topological control of p21^{WAF1/CIP1} expression in normal and neoplastic tissues. *Cancer Res* 1995;55:2910–9.
- Kawamata N, Morosetti R, Miller CW, et al. Molecular analysis of the cyclin-dependent kinase inhibitor gene p27/Kip1 in human malignancies. *Cancer Res* 1995;55:2266–9.
- Kobatake T, Yano M, Toyooka S, et al. Aberrant methylation of p57KIP2 gene in lung and breast cancers and malignant mesotheliomas. *Oncol Rep* 2004;12:1087–92.
- Chen X, Kandasamy K, Srivastava RK. Differential roles of RelA (p65) and c-Rel subunits of nuclear factor κ B in tumor necrosis factor-related apoptosis-inducing ligand signaling. *Cancer Res* 2003;63:1059–66.
- Li F, Ambrosini G, Chu EY, et al. Control of apoptosis and mitotic spindle checkpoint by survivin. *Nature* 1998;396:580–4.
- Xia C, Xu Z, Yuan X, et al. Induction of apoptosis in mesothelioma cells by antisurvivin oligonucleotides. *Mol Cancer Ther* 2002;1:687–94.
- Maeda T, Hanna AN, Sim AB, Chua PP, Chong MT, Tron VA. GADD45 regulates G₂/M arrest, DNA repair, and cell death in keratinocytes following ultraviolet exposure. *J Invest Dermatol* 2002;119:22–6.
- Schmidt M, Fernandez de Mattos S, van der Horst A, et al. Cell cycle inhibition by FoxO forkhead transcription factors involves downregulation of cyclin D. *Mol Cell Biol* 2002;22:7842–52.
- Yakirevich E, Maroun L, Cohen O, Izhak OB, Rennert G, Resnick MB. Apoptosis, proliferation, and Fas (APO-1, CD95)/Fas ligand expression in medullary carcinoma of the breast. *J Pathol* 2000;192:166–73.
- Stahl M, Dijkers PF, Kops GJ, et al. The forkhead transcription factor FoxO regulates transcription of p27Kip1 and Bim in response to IL-2. *J Immunol* 2002;168:5024–31.
- Wonsey DR, Follett MT. Loss of the forkhead transcription factor FoxM1 causes centrosome amplification and mitotic catastrophe. *Cancer Res* 2005;65:5181–9.
- Hirokawa N. Microtubule organization and dynamics dependent on microtubule-associated proteins. *Curr Opin Cell Biol* 1994;6:74–81.
- Bharti AC, Aggarwal BB. Chemopreventive agents induce suppression of nuclear factor- κ B leading to chemosensitization. *Ann N Y Acad Sci* 2002;973:392–5.
- Bharti AC, Aggarwal BB. Nuclear factor- κ B and cancer: its role in prevention and therapy. *Biochem Pharmacol* 2002;64:883–8.
- Clark AS, West K, Streicher S, Dennis PA. Constitutive and inducible Akt activity promotes resistance to chemotherapy, trastuzumab, or tamoxifen in breast cancer cells. *Mol Cancer Ther* 2002;1:707–17.

Therapeutic intervention of experimental breast cancer bone metastasis by Indole-3-carbinol in SCID-hu mouse model

KM Wahidur Rahman*, Fazlul H. Sarkar, Sanjeev Banerjee, Zhiwei Wang, Dezhong J. Liao, Xin Hong and Nurul H. Sarkar¹

Department of Pathology, Karmanos Cancer Institute,
Wayne State University School of Medicine, Detroit, MI and ¹Institute of Molecular Medicine and
Genetics, Medical College of Georgia, Augusta, GA, USA

Running title: Intervention of breast cancer bone metastasis by I3C.

Keywords: I3C, CXCR4, MMP-9, NF- κ B and breast cancer bone metastasis.

*All correspondence to:

KM Wahidur Rahman, Ph.D.

Department of Pathology

Karmanos Cancer Institute

Wayne State University School of Medicine

9374 Scott Hall, 540 East Canfield,

Detroit, MI 48201, USA.

Tel: 313-576-8273

Fax: 313-576-8389

E-mail: kmrahman@med.wayne.edu

ABSTRACT

Several lines of experimental evidence have suggested that chemokine receptor CXCR4, a metastasis promoting molecule, may play important roles in breast cancer bone metastasis. There is emerging evidence linking CXCR4 to MMPs as well as their regulator NF- κ B, a key transcription factor, which is known to activate metastasis-promoting molecules for many types of malignancies including breast cancer. A recent study also showed that promoter region of CXCR4 has several NF- κ B binding sites, suggesting that there may be a cross-talk between CXCR4 and NF- κ B. We have previously shown that Indole-3-carbinol (I3C), a natural compound present in vegetables of the genus *Brassica*, can inhibit NF- κ B in breast cancer cells. However, there are no reports in the literature showing any effect of I3C on CXCR4 expression *in vitro* and *in vivo*. We therefore examined whether I3C could inhibit bone metastasis of breast cancer by inhibiting CXCR4 and MMP-9 expression mediated via the inhibition of the NF- κ B signaling pathway. Here we have modified the SCID-human (SCID-hu) mouse model of experimental bone metastasis for use with the MDA-MB-231 breast cancer cell line. In this animal model, we found that I3C significantly inhibited MDA-MB-231 bone tumor growth, and our results were correlated with the down regulation of NF- κ B. Moreover, we found that I3C significantly inhibited the expression of multiple genes involved in the control of metastasis and invasion *in vitro* and *in vivo*, especially the expression of CXCR4 and MMP-9 along with pro-MMP-9, with concomitant decrease in Bcl-2 and increase in the pro-apoptotic protein, Bax. From these results, we conclude that the CXCR4/NF- κ B pathway is critical during I3C-induced inhibition of experimental breast cancer bone metastasis. These results also suggest that I3C could be a promising agent for the prevention and/or treatment of breast cancer bone metastasis in the future.

INTRODUCTION

Metastasis is a non-random process, and each cancer type has its own preferred sites of metastasis (1, 2). Metastasis of cancer cells is a complex process involving multiple steps including invasion, angiogenesis, intravasation, trafficking of cancer cells through blood vessels, extravasations, organ-specific homing, and growth (3, 4). Bone is one of the most common sites of metastasis for human breast cancer (5). A recent study has shown that a chemokine receptor, CXCR4, is highly expressed in breast cancer cells but not in normal breast tissue while its ligand, stromal derived factor-1 α [SDF-1 α , also called CXCL12], is expressed in those organs where breast cancer metastasis is frequently found (6). Thus, it is quite possible that CXCR4 and/or SDF-1 α signaling may be involved in attracting and homing breast cancer cells in the bone.

There are some protease systems which are required for extracellular matrix degradation for the growth of cancer cells at the metastatic sites. Matrix metalloproteinases (MMPs) are a group of enzymes required for extracellular matrix degradation for the growth of cancer cells at the metastatic sites (3, 4). These enzymes are secreted as pro-proteins and usually need to be activated by urokinase plasminogen activators (3, 4). There is increasing evidence connecting CXCR4 to MMPs, IL-8 and urokinase plasminogen (uPA) (7). MMPs and uPA as well as their regulator NF- κ B, a key transcription factor, are known to activate metastasis-promoting molecules for many types of malignancies including breast cancer (5, 6, 8-13). NF- κ B was also found to be constitutively activated in human breast cancer (14, 15). A recent study and TESS analysis (TRANSFAC 4.0 database version) also showed that the promoter region of CXCR4 has several NF- κ B binding sites, suggesting that there may be a cross-talk between NF- κ B and CXCR4 (16-18). Thus, NF- κ B may regulate the expression and function of CXCR4 which may, in part, be activated via up-regulation of the expression of IL-8, uPA, and MMP-9 (19). Taken together, CXCR4 may be the point of convergence of many mediators of metastatic processes at the metastatic sites.

I3C and its stable condensation product, 3,3'-Diindolylmethane (DIM), are compounds that are abundant in cruciferous vegetables and have been shown to possess inhibitory effects on the growth and metastatic abilities of several prostate and breast cancer cell lines (20-29). It has also been shown that I3C and DIM inhibit cell growth and induce apoptotic cell death by their pleiotropic effects on the regulation of multiple genes such as p21, p27, cyclin-dependent kinase, survivin, Bax/Bcl-2, cytochrome p-450 1A1 and GADD153 (28, 30). Interestingly, our recent

studies have shown that the effect of I3C and DIM is mediated by the inactivation of NF- κ B (23-27, 29). Thus, agents that directly block the expression of CXCR4 signaling, partly due to inhibition of NF- κ B, may have great therapeutic potential for treating metastatic breast cancer. I3C also inhibits the *in vitro* invasive potential of human breast cancer cell lines, suggesting that I3C could inhibit the metastatic growth of breast cancer (31). Collectively, these studies suggest that NF- κ B may play a pivotal role in controlling breast cancer metastasis, a concept supported by many other lines of evidence, in breast cancer and other types of malignancies (23-27, 29). We report here that I3C treatment of MBA-MB-231 breast cancer cells results in decreased CXCR4 mRNA expression. However, the role of I3C in the inhibition of bone metastasis of human breast cancer has not been documented, perhaps due to a lack of appropriate animal models of experimental breast cancer bone metastasis. We hypothesize that I3C-induced inhibition of NF- κ B will inhibit CXCR4 and other NF- κ B targeted genes that could be mechanistically responsible for the inhibition of experimental breast cancer bone metastasis in a suitable animal model.

There are some reports in the literature about the spontaneous animal model of breast cancer (32, 33). A mouse model of human prostate cancer metastasis (SCID-hu) using human prostate cancer cells grown in the human bone implanted into SCID mice has been developed by Nemeth and co-workers (34). But there are no other reports in the literature regarding suitable animal models that could be utilized faithfully for studying human breast cancer bone metastasis using human breast cancer cell lines that grow in the marrow of human bone environment implanted subcutaneously into mice. We believe that the advantage of SCID-hu model may provide a more clinically relevant model for growth of human breast cancer in a human bone microenvironment. For these reasons, here we have adapted the SCID-hu model of experimental bone metastasis for use with the MDA-MB-231 breast cancer cell line, and determined the effect of I3C *in vivo* in this model to test our hypothesis as stated above. We found that I3C could inhibit the growth of MDA-MB-231 cells in a SCID-hu model with concomitant inhibition of CXCR4, MMP-9 and Bcl-2 mediated by the down-regulation of NF- κ B.

MATERIALS AND METHODS

Two-step real-time quantitative reverse transcription-polymerase chain reaction analysis of gene expression in I3C treated MDA-MB-231 breast cancer cells

Total RNA was isolated in I3C treated MDA-MB-231 breast cancer cell line by Trizol (Invitrogen, Carlsbad, CA). One microgram of total RNA was subjected to first strand cDNA synthesis using TaqMan reverse transcription reagents kit (Applied Biosystems, Foster City, CA) in a total volume of 50 microliters including 6.25 U MultiScribe reverse transcriptase and 25 pmol random hexamers. RT reaction was performed with 25°C 10 min, followed by 48°C 30 min and 95°C 5 min. The primers were checked by running a virtual PCR, and primer concentration was optimized to avoid primer dimer formation. Also, dissociation curves were checked to avoid nonspecific amplification. Real-time PCR amplifications were undertaken in Mx4000 Multiplex QPCR System (Stratagene, La Jolla, CA) using 2 × SYBR Green PCR Master Mix (Applied Biosystems). One micro liter RT reaction was used for a total volume of 25 micro liter quantitative PCR reactions. The thermal profile for SYBR real-time PCR was 95 °C 10 min followed by 40 cycles of 95 °C 15 s and 60 °C 1 min.

Animal care and human bone implantation

Female homozygous CB17 scid/scid mice, aged 4 weeks, were purchased from Taconic Farms (Germantown, NY). The mice were maintained according to the National Institutes of Health standards established in the “Guidelines for the Care and Use of Experimental Animals,” and all experimental protocols were approved by the Animal Investigation Committee of Wayne State University (Detroit, MI). Human fetal bone tissue was obtained by a third-party, nonprofit organization (Advanced Bioscience Resources, Alameda, CA), and written informed consent was obtained from the donor, consistent with regulations issued by each state involved and the federal government. Human fetal femurs and humeri of 16–22 weeks of development were divided in half longitudinally and then again in half transversely into four fragments about 1 cm long and 3 or 4 mm in diameter (34). After 1 week of acclimatization, these bone fragments were implanted s.c. in the flank through a small skin incision with the opened marrow cavity against the mouse muscle. Isoflurane anesthesia was used during all surgical procedures.

Production of breast cancer bone tumors and I3C treatment

Suspensions of MDA-MB-231 cells (2×10^5 cells in a volume of 20 μ l of PBS) were injected using a 27-gauge needle through the mouse skin directly into the marrow of implanted fetal bone. The mice were then divided into two groups: Control (n = 10) and Intervention (n = 10) groups (Fig 2). Sesame seed oil was used to facilitate gavage and avoid irritation of the esophagus and was safe as shown also by others (35, 36). The mice in the intervention group were given I3C (1 mg/day/mouse) by oral gavage everyday for five weeks as soon as the majority of the bone implants began to enlarge (now called a “bone tumor”) as determined by caliper measurements (23rd day after cancer cell injection). The control mice received only sesame seed oil without I3C. The volume of the bone tumor in each group was determined by twice-weekly caliper measurements according to the formula $ab^2 / 2$, where a = length and b = shortest measurement. Percent (%) reduction in tumor volume at the end of the treatment was deduced by the formula: volume of tumor in experimental group \div volume in control mice, $\times 100$. The statistical significance of differential findings between the experimental groups and control was determined by student *t*-test as implemented by Excel 2000 (Microsoft Corp., Redmond, WA). The mice were sacrificed 3 months after cell injection. Bone tumors were subjected to *ex vivo* imaging on a Lo-Rad M-IV mammography unit (Karmanos Cancer Institute, Detroit, MI) using a magnified specimen technique. Images were developed using a Kodak 2000 screen and radiography film (Kodak, Rochester, NY). Upon sacrifice, tumor tissue from each mouse was harvested and cut into two pieces; one part was frozen for molecular analysis, and the other part of the tissue was fixed in formalin and embedded in paraffin for histological evaluation and immunohistochemistry.

Tissue collection, fixation and H&E staining

Freshly harvested tumors grown in the implanted bones were fixed in 10% buffered formalin for 48 hours and decalcified with 10% EDTA, embedded and sectioned. Samples were then washed with tap water and soaked in a graded series of 50%, 60%, 70%, 80% and 90% ethanol for 30 minutes, and then in 90% and 100% ethanol for one hour. They were then held in a solution of 100% ethanol and xylene at a 1:1 ratio for 30 minutes before being embedded in paraffin and held at 60°C for one hour to make paraffin blocks. Transverse sections (5 μ m) were taken from the blocks and prepared for histochemical and immunohistochemical staining. Haematoxylin & Eosin (H&E) staining was used for histological observation.

Electrophoretic mobility shift assay for measuring NF- κ B activity

MDA-MB-231 cells were plated at a density of 1×10^6 cells in 100 mm dishes and cultured for 24 hours. Subsequently, the cultures were treated with 60 and 100 μ M I3C or DMSO for 24, 48 and 72 hours. I3C (Sigma) was dissolved in dimethylsulfoxide (DMSO; final concentration 0.1%) to make a 10 mM stock solution and was added directly to the culture media at different concentrations. Nuclear extracts were prepared from control and I3C-treated breast epithelial cells as previously described (25, 29, 37) and subjected to analysis for NF- κ B DNA binding activity as measured by electrophoretic mobility shift assay. Using frozen tumor tissue, nuclear proteins were also extracted as previously described (25, 29, 37). Briefly, tissues were minced and incubated on ice for 30 min in 0.5 ml of ice-cold buffer A composed of 10 mM HEPES (pH 7.9), 1.5 mM KCl, 10 mM $MgCl_2$, 0.5 mM DTT, 10% NP-40, 0.1% IGEPAL CA-630 and 0.5 mM PMSF. The minced tissue was homogenized using a Dounce homogenizer (Kontes Co., Vineland, NJ) followed by centrifugation at $5000 \times g$ at $4^\circ C$ for 10 min. The supernatant (cytosolic proteins) was collected for Western Blot analysis and kept at $-70^\circ C$ until use. The crude nuclear pellet was suspended in 200 μ l of buffer B (20 mM HEPES, pH 7.9; 25% glycerol, 1.5 mM $MgCl_2$; 420 mM NaCl; 0.5 mM DTT; 0.2 mM EDTA; 0.5 mM PMSF; and 4 μ M leupeptidin) and incubated on ice for 30 min. The suspension was centrifuged at $16,000 \times g$ at $4^\circ C$ for 30 min. The supernatant (nuclear proteins) was collected and kept at $-70^\circ C$ until use. The protein concentration was determined using BCA protein assay (Pierce Chemical Co., Rockford, IL). Electrophoretic mobility shift assay was done by pre-incubating 8.0 μ g of nuclear extract with a binding buffer containing 20% glycerol, 100 mM $MgCl_2$, 2.5 mmol/L EDTA, 2.5 mmol/L DTT, 250 mmol/L NaCl, 50 mmol/L Tris-HCl, and 0.25 mg/mL poly (dI:dC) for 10 minutes. After the addition of IRDye-700 labeled NF- κ B oligonucleotide, samples were incubated for an additional 20 minutes. The DNA-protein complexes were electrophoresed in an 8.0% native polyacrylamide gel and then visualized by Odyssey Infrared Imaging System using Odyssey Software Release 1.1.

Western Blot analysis for measuring the protein levels of Bcl-2, Bax, CXCR4 and MMP-9 activity

MDA-MB-231 cells were plated on culture dishes and allowed to attach for 24 hours followed by the addition of 60 or 100 μ M I3C and incubated for 24, 48, and 72 hours. Control cells were incubated in the medium with DMSO for similar times. Total cell lysates were prepared using the method as previously described (25, 29, 37). Using frozen tumor tissue, nuclear proteins were

extracted using the method as previously described (25, 29, 37). Briefly, tissues were minced and incubated on ice for 30 min in 0.5 ml of ice-cold buffer A, composed of 10 mM HEPES (pH 7.9), 1.5 mM KCl, 10 mM MgCl₂, 0.5 mM DTT, 10% NP-40, 0.1% IGEPAL CA-630 and 0.5 mM PMSF. The minced tissue was homogenized using a Dounce homogenizer (Kontes Co., Vineland, NJ) followed by centrifuging at 5000 x g at 4°C for 10 min. The supernatant (cytosolic proteins) was collected for Western Blot analysis. Protein content was quantified with a BCA protein assay (Pierce, Rockford, IL), and equal amounts of proteins were resolved by 10, 12 and 14% SDS PAGE. Immunoblot was performed with antibodies to Bcl-2 (Calbiochem, CA), Bax and CXCR4 (Santa Cruz Biotechnology, Inc., CA), MMP-9 (Santa Cruz Biotechnology, Inc., CA) and β -actin (Sigma-Aldrich). A representative blot from three independent experiments was presented.

Immunohistochemical staining for CXCR4 and MMP-9

Freshly harvested tumors grown in the implanted bones were fixed in 10% buffered formalin, decalcified, embedded, and sectioned. The paraffin sections of tumor tissues were deparaffinized then rehydrated through a graded alcohol series. Slides were placed in 10 mM citrate buffer (pH 6.0) and heated in a microwave for 3 minutes. Non-specific sites were blocked by incubation with Superbloc (ScyTek, Logan, UT). Sections were incubated with antibodies to MMP-9 (Calbiochem, CA) and CXCR4 (Santa Cruz Biotechnology, Inc., CA) followed by staining with appropriate HRP-conjugated secondary antibodies followed by color development. The stained slides were dehydrated and mounted in per mount and visualized using an Olympus microscope (Olympus, Japan). Images were captured with an attached camera linked to a computer.

MMP-9 activity assay

MDA-MB-231 cells were seeded in a 6 well plate (1.0×10^5 cells per well) and incubated at 37°C. After 24 hours, the complete medium was removed and the cells were washed with serum-free medium. The cells were then incubated in serum-free medium supplemented with 60 or 100 μ M I3C for 72 hours. MMP-9 activity in the conditioned medium and cell lysate was detected by using Fluorokine E Human MMP-9 Activity Assay Kit (R&D Systems) according to the manufacturer's protocol.

Invasion assay

The invasive activity of MDA-MB-231 cells with different treatments was tested by using BD BioCoat Tumor Invasion Assay System (BD Biosciences, Bedford, MA) according to the manufacturer's protocol with minor modification. Briefly, MDA-MB-231 cells (5×10^4) with serum free medium supplemented with 60 or 100 μ M I3C were seeded into the upper chamber of the system. Bottom wells in the system were filled with complete medium and the same reagent treatment as the upper chamber. After 48 hours of incubation, the cells in the upper chamber were removed, and the cells, which invaded through the matrigel matrix membrane, were stained with 4 μ g/ml Calcein AM in Hanks buffered saline at 37°C for one hour. Then, fluorescence of the invaded cells was read in ULTRA Multifunctional Microplate Reader (TECAN) at excitation/emission wavelengths of 530/590 nm. These fluorescently labeled invasive cells were also photographed under a fluorescent microscope.

Statistical Analysis

The statistical significance was determined using student's *t*-test, and $P < 0.05$ was considered significant.

RESULTS

Regulation of CXCR4 mRNA expression by I3C treatment

A recent study has indicated that the MDA-MB-231 cell line highly expresses CXCR4 (17), and I3C has been shown to possess inhibitory effects on the growth and metastatic abilities of several cancers including breast cancer (24-27, 29, 38-43). In order to confirm the alternations of CXCR4 expression after I3C treatment in MDA-MB-231 breast cancer cells, we conducted real-time RT-PCR analysis for the CXCR4 gene (Fig 1). The altered mRNA expression of CXCR4 was observed as early as 24 h after I3C treatment and was significantly more evident after 72 hours treatment. The results of RT-PCR analysis for CXCR4 gene along with published *in vitro* results suggest that I3C could regulate the transcription of genes involved in angiogenesis, tumor cell invasion and metastasis (27-30, 38, 44-46). However, we asked the most important question whether I3C could affect tumor growth in the bone environment, which is answered by the following experiments.

Inhibition of bone tumor growth by I3C

We report here, for the first time, use of the SCID-hu animal model (34) as an experimental model for breast cancer bone metastasis and the effect of I3C *in vivo* in impeding tumor growth and metastasis. The experimental design and treatment schedule is depicted in fig 2. Currently there are no reports in the literature documenting any impact of the host (i.e., mouse) environment on the bone xenograft (34, 47). Under our experimental conditions, administration of I3C by gavage treatment caused 50% reduction in tumor volume (Fig 3A), compared to control group. The statistical analysis indicated that compared to the control group, bone tumor growth was significantly lower in the intervention ($P<0.05$) group. The experiment was terminated 3 months after I3C treatment. We found that I3C (1 mg I3C/day/mouse) significantly inhibited breast cancer bone tumor growth (Fig 3A and 3B) and osteolysis, documenting the efficacy of I3C in inhibiting breast cancer cell growth in the experimental model of bone metastasis. *Ex vivo* bone tumor X-ray showing bone osteolysis and tumor growth of MDA-MB-231 cells in the control and I3C treatment group. Arrow indicates less residual bone in the control group (3B I) relative to I3C treated group (3B II). We found that treatment with I3C significantly inhibited cellular growth of breast cancer cells in a bone environment and also inhibited bone osteolysis in SCID-hu mice, demonstrating an inhibitory effect of I3C in an *in vivo* model of experimental breast cancer bone metastasis. At

autopsy, all tumors were found localized at the site of injection with essentially spread to no other organs. Our treatment conditions did not cause any weight loss of the animals, suggesting that I3C did not induce any deleterious effects under the present experimental conditions. These results illustrate, for the first time, the efficacy of I3C in inhibiting tumor growth in an experimental breast cancer bone metastasis model.

Tumor histology and inhibition of osteolysis by I3C

H&E histology evaluation showed typical osteolytic bone metastasis of MDA-MB-231 cells in SCID-hu animals (Fig 3C). Human fetal bones subcutaneously implanted in SCID, mice as shown by representative figure, were randomly selected from each group (Fig 3C I). Suspensions of MDA-MB-231 cells (2×10^5 cells in a volume of 20 μ l of PBS) were injected using a 27-gauge needle through the mouse skin directly into the marrow of implanted fetal bone. Black arrow represents the site of significant progression of tumor growth as shown by representative figure from untreated group (Fig 3 C II). Representative figure of mice from treatment group photographed at the time of sacrifice. Red arrow represents the site of significant regression of tumor growth in I3C treatment group (Fig 3C III). In the control group, continuous osteolytic and invasive growth into the adjacent bone resulted in gradual loss of the bone structure (Fig 3C IV, black arrow). Yellow arrows indicate residual bone surrounded by invasive tumor cells (Fig 3C V). In contrast, the group receiving I3C treatment showed pronounced death of tumor cells (yellow arrows) and preservation of the bone structure (black arrow head) (Fig 3C VI). In order to further demonstrate whether the anti-tumor effect observed by I3C treatment could be due to inactivation of NF- κ B, we performed the following experiments.

Inhibition of NF- κ B activation by I3C

To answer the most important question, whether treatment of animals with I3C could effectively target a specific signaling molecule such as NF- κ B in tumor tissues, nuclear extract from frozen tumor tissues (randomly selected from control and treatment groups) was subjected to analysis for NF- κ B DNA binding activity as measured by electrophoretic mobility shift assay. The results are shown in Fig 4B, which clearly show that I3C was effective in down-regulating NF- κ B DNA binding activity in animal tumors receiving I3C compared to control tumors. These *in vivo* results were similar to our *in vitro* findings (Fig 4A), including our previously published data (27, 29), suggesting that the inactivation of NF- κ B is at least one of the molecular mechanisms by which

I3C induced anti-tumor activity in our experimental animal model. The specificity of NF- κ B DNA binding activity was confirmed by supershift assays (Fig 4C). Retinoblastoma (Rb) protein level served as nuclear protein loading control. These data also provide proof of principle and suggest that I3C could be an effective anti-tumor agent in an animal model of breast cancer bone metastasis, which is mediated by inactivation of NF- κ B DNA binding activity (27, 29). The inactivation of NF- κ B may decrease the expression of NF- κ B downstream genes such as CXCR4 and Bcl-2 proteins with concomitant increase in the pro-apoptotic protein Bax in addition to the alterations in the expression of genes which are known to be involved in the osteolytic and apoptotic processes.

Effect of I3C on the expression of Bcl-2, Bax, CXCR4 and MMP-9

The effect of CXCR4 expression in tumor tissue (tissue randomly selected from control and treatment groups) was studied by Western Blot analysis, and we found a significant down-regulation in the expression of CXCR4 and MMP-9 in tumors of the I3C-treated group (Fig 5B, lane 2), compared to the untreated group (Fig 5B, lane 1). These *in vivo* results were similar to our *in vitro* findings (Fig 5A, lane 2 and 3). It is known that CXCR4 protein family significantly increases the viability of CD34(+) T-cell precursors by modulating the expression of Bcl-2 and Bax genes and consequently stimulating the proliferation of CD34(+) thymic precursor cells (48). We found that I3C caused a decrease in the expression of Bcl-2 and an increase in the expression of Bax in I3C treated tumors (Fig 5B) and MDA-MB-231-cells (Fig 5A). These results are consistent with our previously published *in vitro* data on Bax and Bcl-2 expression in cells treated with I3C (24, 25). Moreover, recent evidence indicates that deregulation of MMP expression contributes to invasion, metastasis, and tumor angiogenesis (49-55). For example, over-expression of MMP-2 and MMP-9 has been reported to be positively associated with the progression of human cancers (49-58). Our present results suggest that I3C can inhibit metastasis of breast cancer cells mediated by the inhibition of MMP-9 expression. These results also provide evidence in support of our hypothesis, implicating the role of CXCR4 in metastatic tumor cells by controlling tumor cell migration. Remarkably, we show that CXCR4 may protect tumor cells by reducing apoptotic processes, resulting in increased tumor cell growth and viability. This protective effect involves both the down-regulation of the pro-apoptotic Bax protein and the up-regulation of the anti-apoptotic Bcl-2 protein. The level of Bcl-2 expression was found to be significantly down-regulated in I3C treated animal tumors and MDA-MB-231 cells (Fig 5A and 5B). In contrast, the expression of Bax was found to be significantly up-regulated in I3C treated animals and MDA-MB-231 cells

(Fig 5A and 5B). Collectively, our results implicate the role of CXCR4/NF- κ B pathway as a critical event during I3C-induced apoptosis and inhibition of tumor cell proliferation. These preclinical data suggest that I3C could have beneficial effects in patients with breast cancer bone metastasis.

Analysis of CXCR4 and MMP-9 expression by immunohistochemistry

Previous studies have shown over-expression of chemokine receptor CXCR4 and/or SDF1- α in metastatic breast cancer cells (6), and these chemokine protein families are also known to activate other metastasis-promoting molecules, such as MMPs, in breast cancer cells (17). Thus, agents that directly block the expression of CXCR-4 could also down-regulate MMP-9 expression which could also be partly due to inactivation of NF- κ B, and this could have great therapeutic potential for treating metastatic breast cancer. In order to verify the alteration of CXCR4 and MMP-9 by I3C treatment, we conducted immunohistochemical analysis for CXCR4 and MMP-9 protein. The expression of CXCR4 (Fig 6 upper panel) and MMP-9 (Fig 6 lower panel), two important molecules of tumor cell survival and metastasis that are downstream of NF- κ B, was significantly decreased in MDA-MB-231 bone tumors in SCID-Hu mice receiving the I3C (Fig 6B and 6D) compared to control (Fig 6A and 6C). Overall, our results suggest that the inactivation of NF- κ B may indeed reduce the levels of CXCR4 and MMP-9 in the tumor microenvironment resulting in anti-tumor activity of I3C in our experimental model of breast cancer bone metastasis.

Regulation of the expression and activity of MMP-9 by I3C

Treatment with I3C showed down-regulation of MMP-9 by Western blot analysis. Furthermore, we found that I3C significantly inhibited the activity of MMP-9 in conditioned medium in MDA-MB-231 cell culture (Fig. 7A). Together, these results clearly suggest that I3C down-regulates MMP-9, and thus I3C could enhance the anti-tumor and anti-metastatic activity in MDA-MB-231 breast cancer cells. In order to further support the role of MMP in breast cancer metastasis, we conducted cell invasion assay.

I3C inhibits cancer cell invasion

It has been well known that MMP-9 is an important molecule involved in cancer cell invasion and metastasis. Because I3C inhibited the expression and activity of MMP-9, we tested the effects of I3C on cancer cell invasion. We found that I3C inhibited invasion of MDA-MB-231 cells through matrigel matrix membrane compared to control (Fig 7B and 7C).

DISCUSSION

The importance of this study lies in the use of a SCID-hu mouse model system that resembles human disease and may also be more relevant to clinical breast cancer than other animal models. Typical osteolytic bone metastasis of MDA-MB-231 cells in a SCID-hu model showed continuous osteolytic and invasive processes resulting in the loss of the bone structure and increased tumor growth, which are believed to be associated with activation of CXCR4/NF- κ B signaling pathways (6, 17), similar to those observed in many human breast cancers (6, 17, 59). Our present observations are also in direct agreement with reports showing a significant elevation of CXCR4 in models of metastatic breast cancer with immunodeficient mice (60). Thus, our SCID-hu animal model of breast cancer bone metastasis has offered us an experimental model in which we have tested the effects of I3C as a chemopreventive or therapeutic agent against breast cancer.

I3C has been shown to target multiple pathways of tumorigenesis including proliferation, apoptosis, angiogenesis, invasion, and tumor-induced immune suppression in various breast tumor cell lines (23-27, 29, 38-43, 61-64). Several studies have suggested that chemokine receptor CXCR4 is over-expressed in metastatic breast cancer cells (6). However, no reports exist in the literature elaborating how I3C inhibits CXCR4, a metastatic promoting molecule *in vivo* model of breast cancer cells. In the current report, we show, for the first time, that I3C is an effective agent in the inhibition of cellular growth of breast cancer in a bone environment using an animal model of experimental breast cancer bone metastasis. The anti-tumor activity of I3C was regulated by the inhibition of CXCR4, which is known to regulate the organ-specific trafficking and invasion of metastatic tumor cells mediated by inactivation of NF- κ B.

Chemokines and chemokine receptors are believed to play important roles in metastatic breast cancer (6, 59). The most common sites of breast cancer metastases include the lung, liver, lymph node, and bone in which CXCR4 was found to be highly expressed (6). Moreover, NF- κ B, a key transcription factor, may regulate the expression and function of CXCR4 which in turn may activate other metastasis-promoting molecules such as MMP-9 in breast cancer cells (17). A recent study also showed that the promoter region of CXCR4 has several NF- κ B binding sites, suggesting that there may be a cross-talk between NF- κ B and CXCR4 (17). In models of metastatic breast cancer, the administration of neutralizing antibodies or peptide antagonists of CXCR4 substantially reduced lung metastases (6, 65). CXCR4 expression was found to be up-regulated in murine lung metastases in a NF- κ B-dependent fashion (17), again suggesting that CXCR4 signaling is an

important determinant of metastatic breast cancer. Moreover, a significant correlation between the relative expression levels of CXCR4 and the extent of lymph node metastases was found in invasive ductal carcinomas of the breast in humans. The expression of CXCR4 was also found to be increased in invasive ductal carcinoma specimens, compared with ductal carcinoma *in situ* and normal breast epithelium. CXCR4 expression by normal breast stromal cells adjacent to malignant cells was also increased (66). We anticipated that CXCR4 in the tumor microenvironment may function to promote breast cancer proliferation, migration, and invasion, and our current data suggest that I3C could interrupt this signaling pathway, resulting in tumor growth inhibition of breast cancer bone metastasis.

Our present data recapitulates recent *in vitro* results in breast cancer cell lines, showing that I3C induces apoptosis by blocking NF- κ B activation (27, 29). Moreover, NF- κ B, a key transcription factor, may regulate the expression and function of CXCR4 and activate other metastasis-promoting molecules (uPA, MMPs) in breast cancer cells (17). Decreased expression of CXCR4 was associated with decreased expression of NF- κ B which in turn reduced the levels of MMP-9 in tumors treated with I3C. This could be one of the mechanism(s) underlying enhanced tumor cell apoptosis and reduced tumor cell proliferation *in vivo*. Taken together, our results suggest multiple functions of NF- κ B in the growth, migration, and organ-specific metastasis of breast cancer cells which, in part, appear to be mediated through the induction of CXCR4. Moreover, the CXCR4/NF- κ B pathways activated MMP-9 expression in breast cancer bone metastasis and were also abrogated by I3C, thus suggesting that our model could be useful to fully evaluate the role of CXCR4/NF- κ B pathways *in vivo* as well as its mechanistic role in the inhibition of tumor cell invasion and metastasis (Fig 8).

Several other potential mechanism(s) could also explain the regulation of CXCR4 associated with breast cancer metastasis (Fig 8). One potential mechanism that has been associated with CXCR4-related inhibition of apoptosis is that CXCR4 reduces the apoptotic rate by increasing the level Bcl-2 expression and decreasing the level of Bax expression (48). Our *in vivo* and *in vitro* data support this concept because inhibiting NF- κ B production by targeting CXCR4 activity in the SCID-hu tumors led to a decrease in Bcl-2 protein levels, a concomitant increase in the pro-apoptotic protein Bax and decreased MMP-9 expression. These results are consistent with our previously published *in vitro* data on Bax/Bcl-2 expression (24, 25) and provide support for our hypothesis, implying the role of CXCR4 in metastatic tumor cells, suggesting that I3C could be beneficial in patients with breast cancer bone metastasis.

In addition to these mechanism(s), MMP-9 has also been reported to be involved in cancer invasion and metastasis (52, 53, 55). Several studies have shown that MMPs plays prominent roles in metastasis (49-53, 55). MMP-9 has been implicated in metastasis because of its role in the degradation of basement membrane collagen. In addition, MMP activity is known to play a role in both normal and cancer-induced bone remodeling. It has been reported that NF- κ B and Matrix metalloproteinases (MMPs) are involved in the processes of tumor invasion and metastasis (67). In this study, we found that I3C inhibited the expression, secretion, and activation of MMP-9, suggesting that I3C could prevent bone matrix degradation and reduce breast cancer cell growth in human bone implanted in SCID mice. The down-regulation of MMP-9 by I3C could be mediated by the down-regulation of NF- κ B whose binding site has been found in the promoter of MMP-9 (67, 68). These results suggest that I3C could promote the anti-tumor and anti-metastasis activities in SCID-hu model of breast cancer bone metastasis partly through the down-regulation of MMP expression (Fig 8). Since we observed that I3C down-regulated MMP-9, we tested the effects of I3C on the invasion of MDA-MB-231 breast cancer cells and found that I3C inhibited the invasion of MDA-MB-231 breast cancer cells. These results corresponded with MMP-9 data, showing that I3C could inhibit cancer cell invasion partly through down-regulation of MMP-9.

In summary, I3C treatment may exert its anti-proliferative, anti-angiogenic, and pro-apoptotic effects by decreasing CXCR4 and MMP-9 expression which was associated via the down-regulation of the NF- κ B pathway. In this report, we have shown that reduced expression of CXCR4 by I3C limits cellular growth of breast cancer cells in a bone environment *and* prevents breast cancer bone metastasis. These results further extend the potential therapeutic application of I3C for metastatic breast cancer.

ACKNOWLEDGEMENTS

This work was supported by Concept Awards funded by Department of Defense (DOD) to KM Wahidur Rahman. Grant number: W81XWH-04-1-0689 and W81XWH-05-1-0505. We thank Carrie Koerner for her editorial assistance.

FIGURE LEGENDS:

Fig 1: Real-time RT-PCR analysis of CXCR4 genes. Comparative analysis shows the down-regulation of CXCR4 mRNA expression in MDA-MB-231 cells grown in culture after 72 hour treatment with I3C compared to control. *, $P < 0.05$

Fig 2: Flow chart representation of *in vivo* experimental design and treatment schedule. The human fetal bone was implanted subcutaneously in the right flank of female homozygous CB17 SCID mice through a small skin incision with the open marrow cavity against the mouse muscle. Four weeks after the bone implantation, suspensions of MDA-MB-231 cells (2×10^5 cells) were injected through the mouse skin directly into the marrow of implanted fetal bone. The mice in the intervention group were given I3C (1 mg/day/mouse) by oral gavage everyday for five weeks as soon as the majority of the bone implants began to enlarge (now called a “bone tumor”) as determined by caliper measurements (23rd day after cancer cell injection). The control mice received only saline without I3C. The experiment was terminated 3 months after I3C treatment.

Fig 3A: Inhibitory effects of I3C on the growth of xenograft bone tumors of MDA-MB-231 cells in SCID-hu mice. A total of 20 mice were divided into 2 groups. Under the experimental conditions, administration of I3C by gavage treatment caused 50% reduction in tumor volume, compared to control group. *, $P < 0.05$

B. *Ex vivo* bone tumor X-ray showing bone osteolysis and tumor growth of MDA-MB-231 cells in the control and I3C treated group. Arrow indicates less residual bone in the control group (B I) relative to I3C treated group (B II). We found that treatment with I3C inhibited tumor growth and osteolytic bone destruction resulting in more residual bone volume compared to control group (B I).

C. Typical osteolytic bone metastasis of MDA-MB-231 cells in SCID-hu model. (C I): Human fetal bone subcutaneously implanted in SCID mice; (C II): MDA-MB-231 cells (2×10^5 cells) were injected directly into the marrow of implanted bone. Black arrow represents the site of significant progression of tumor growth in representative untreated control mice; (C III): Representative figure of mice from treatment group photographed at time of sacrifice showing significant regression of

tumor growth (red arrow) by I3C treatment; (C IV): MDA-MB-231 cells were injected into the implanted human fetal bone showing invasion (black arrow); (C V): Continuous osteolytic and invasive process resulting in gradual loss of the bone structure. Yellow arrows indicate residual bone surrounded by invasive tumor cells; (C VI): Histological evaluation showing I3C treatment resulted in pronounced death of tumor cells (yellow arrows) and preservation of the bone structure (black arrow head).

Fig 4: I3C abrogates NF- κ B DNA binding activity.

A. I3C abrogates NF- κ B DNA binding activity in MDA-MB-231 breast cancer cells. Cells were treated with 60 or 100 μ M I3C (Lane 2 and 3, respectively) for 72 hours. Nuclear extracts were prepared from control and I3C-treated cells and subjected to analysis for NF- κ B DNA binding activity as measured by electrophoretic mobility shift assay.

B. MDA-MB-231 cells were grown in the marrow surface of the previously implanted bone in SCID animal. After the termination, tumors were removed and the nuclear protein were subjected to analysis the DNA binding activity of NF- κ B as measured by electrophoretic mobility shift assay (EMSA). Gel shift assay for NF- κ B done on randomly selected frozen tumor tissues obtained from each treatment groups of animals. Group-1 (Lane 1, control animal; Lane 2 and 3, animal received I3C). Results showed that I3C was effective in down-regulating NF- κ B in treated animals relative to control tumors. Arrow indicates NF- κ B DNA binding activity. **C.** The specificity of NF- κ B DNA binding activity was confirmed by supershift assays. Retinoblastoma (Rb) protein level served as nuclear protein loading control.

Fig 5: I3C-induced alterations in metastasis promoting and apoptosis related genes.

A. I3C down-regulated metastasis related protein such as Bcl-2, Bax, CXCR4 and MMP-9 expression in total cell lysate. Control, MDA-MB-231 cells treated with DMSO; Day 3, cells treated with 60 or 100 μ M I3C (Lane 2 and 3) for 72 hours. Whole cell lysates were prepared and proteins were subjected to Western blot analysis.

B. Western blot analysis was done on randomly selected frozen tumor tissues obtained from each treatment group of animals. Group-1 (Lane 1, control animal; Lane 2, animal received I3C). Results from Western blot analysis showed that the levels of Bcl-2, Bax, CXCR4 and MMP-9 were significantly down-regulated in I3C treated MDA-MB-231 cells and in animal tumors compared to control group.

Fig 6: Immunohistochemical staining for CXCR4 and MMP-9 in I3C treated and untreated SCID-Hu model and animal tumors done on randomly selected tumor tissues. Tumor cells in untreated control group show intensive staining of CXCR4 (A) and MMP-9 (C). In the contrast, tumor cells in I3C treated SCID-Hu mice show much weaker staining of CXCR4 (B) and MMP-9 (D). Results from immunohistochemical staining showed that the expression of CXCR4 and MMP-9 was significantly decreased in MDA-MB-231 bone tumors in SCID-hu mice receiving the I3C-containing diet.

Fig 7: **A:** MMP-9 expression was down-regulated by I3C. MMP-9 activity assay showed that MMP-9 was down-regulated by I3C in the conditioned medium of MDA-MB-231 breast cancer cells. **B:** I3C induced inhibition of MDA-MB-231 cell invasion. Invasion assay showed that I3C significantly inhibited the invasion of MDA-MB-231 cells through matrigel matrix membrane. **C:** The graphs show the value of fluorescence from the invaded MDA-MB-231 breast cancer cells. The value indicates the comparative amount of invaded MDA-MB-231 breast cancer cells. *, $P < 0.05$; **, $P < 0.01$.

Fig 8: Hypothetical diagram showing the role of CXCR4, MMP-9 and/or other genes in breast cancer cells in a metastatic model and further showing that these molecules could be inhibited by I3C mediated by the inactivation of the NF- κ B pathway.

REFERENCES:

1. Liotta LA. An attractive force in metastasis. *Nature* 2001;410:24-25.
2. Liotta LA, Kohn EC. The microenvironment of the tumour-host interface. *Nature* 2001;411:375-379.
3. Chambers AF, Matrisian LM. Changing views of the role of matrix metalloproteinases in metastasis. *J Natl Cancer Inst* 1997;89:1260-1270.
4. Edward KL, jeffery RM. Estrogen-induced mitogenesis of MCF-7 cells does not require the induction of mitogen-activated protein kinase activity. *J. Steroid Biochem. Mol. Biol.* 2000;75:11-20.
5. Coleman RE, Rubens RD. The clinical course of bone metastases from breast cancer. *Br J Cancer* 1987;55:61-66.
6. Muller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis. *Nature* 2001;410:50-56.
7. Chinni SR, Sivalogan S, Dong Z, et al. CXCL12/CXCR4 signaling activates Akt-1 and MMP-9 expression in prostate cancer cells: the role of bone microenvironment-associated CXCL12. *Prostate* 2006;66:32-48.
8. Han YP, Hughes MW, Nien YD, et al. IL-8-stimulated expression of urokinase-type plasminogen activator in human skin and human epidermal cells. *J Surg Res* 2002;106:328-334.
9. Lane WJ, Dias S, Hattori K, et al. Stromal-derived factor 1-induced megakaryocyte migration and platelet production is dependent on matrix metalloproteinases. *Blood* 2000;96:4152-4159.
10. Moore MA. The role of chemoattraction in cancer metastases. *Bioessays* 2001;23:674-676.
11. Murphy P. Chemokines and the molecular basis of cancer metastasis. *N.Engl. J. Med.* 2001;345:833-835.
12. Nakshatri H, Goulet RJ, Jr. NF-kappaB and breast cancer. *Curr Probl Cancer* 2002;26:282-309.
13. Taichman RS, Cooper C, Keller ET, et al. Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res* 2002;62:1832-1837.
14. Pallares J, Martinez-Guitarte JL, Dolcet X, et al. Abnormalities in the NF-kappaB family and related proteins in endometrial carcinoma. *J Pathol* 2004;204:569-577.

15. Wu JT, Kral JG. The NF-kappaB/IkappaB signaling system: a molecular target in breast cancer therapy. *J Surg Res* 2005;123:158-169.
16. Ganju RK, Brubaker SA, Meyer J, et al. The alpha-chemokine, stromal cell-derived factor-1alpha, binds to the transmembrane G-protein-coupled CXCR-4 receptor and activates multiple signal transduction pathways. *J Biol Chem* 1998;273:23169-23175.
17. Helbig G, Christopherson KW, 2nd, Bhat-Nakshatri P, et al. NF-kappa B promotes breast cancer cell migration and metastasis by inducing the expression of the chemokine receptor CXCR4. *J Biol Chem* 2003
18. Kukreja P, Abdel-Mageed AB, Mondal D, et al. Up-regulation of CXCR4 expression in PC-3 cells by stromal-derived factor-1alpha (CXCL12) increases endothelial adhesion and transendothelial migration: role of MEK/ERK signaling pathway-dependent NF-kappaB activation. *Cancer Res* 2005;65:9891-9898.
19. Karin M, Cao Y, Greten FR, et al. NF-kappaB in cancer: from innocent bystander to major culprit. *Nat Rev Cancer* 2002;2:301-310.
20. Chinni SR, Li Y, Upadhyay S, et al. Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene* 2001;20:2927-2936.
21. Chinni SR, Sarkar FH. Akt inactivation is a key event in indole-3-carbinol-induced apoptosis in PC-3 cells. *Clin Cancer Res* 2002;8:1228-1236.
22. Li Y, Chinni SR, Sarkar FH. Selective growth regulatory and pro-apoptotic effects of DIM is mediated by AKT and NF-kappaB pathways in prostate cancer cells. *Front Biosci* 2005;10:236-243.
23. Li Y, Li X, Sarkar FH. Gene Expression Profiles of I3C- and DIM-Treated PC3 Human Prostate Cancer Cells Determined by cDNA Microarray Analysis. *J Nutr* 2003;133:1011-1019.
24. Rahman KM, Aranha O, Glazyrin A, et al. Translocation of Bax to mitochondria induces apoptotic cell death in indole-3-carbinol (I3C) treated breast cancer cells. *Oncogene* 2000;19:5764-5771.
25. Rahman KM, Aranha O, Sarkar FH. Indole-3-carbinol (I3C) induces apoptosis in tumorigenic but not in nontumorigenic breast epithelial cells. *Nutr Cancer* 2003;45:101-112.
26. Rahman KM, Sarkar FH. Steroid hormone mimics: molecular mechanisms of cell growth and apoptosis in normal and malignant mammary epithelial cells. *J Steroid Biochem Mol Biol* 2002;80:191-201.

27. Rahman KW, Li Y, Sarkar F. Inactivation of Akt and NF- κ B plays important roles during I3c-induced Apoptosis in Breast Cancer cells. *Nutrition and Cancer* 2004;48:84-94.
28. Rahman KW, Li Y, Wang Z, et al. Gene expression profiling revealed survivin as a target of DIM-induced cell growth inhibition and apoptosis in breast cancer cells. *Cancer Research* 2006;In press:
29. Rahman KW, Sarkar FH. Inhibition of nuclear translocation of nuclear factor- κ B contributes to 3,3'-diindolylmethane-induced apoptosis in breast cancer cells. *Cancer Res* 2005;65:364-371.
30. Kim YS, Milner JA. Targets for indole-3-carbinol in cancer prevention. *J Nutr Biochem* 2005;16:65-73.
31. Meng F, Liu L, Chin PC, et al. Akt is a downstream target of NF- κ B. *J Biol Chem* 2002;277:29674-29680.
32. Basu GD, Pathangey LB, Tinder TL, et al. Cyclooxygenase-2 inhibitor induces apoptosis in breast cancer cells in an in vivo model of spontaneous metastatic breast cancer. *Mol Cancer Res* 2004;2:632-642.
33. Eckhardt BL, Parker BS, van Laar RK, et al. Genomic analysis of a spontaneous model of breast cancer metastasis to bone reveals a role for the extracellular matrix. *Mol Cancer Res* 2005;3:1-13.
34. Nemeth JA, Harb JF, Barroso U, Jr., et al. Severe combined immunodeficient-hu model of human prostate cancer metastasis to human bone. *Cancer Res* 1999;59:1987-1993.
35. Hillman GG, Wang Y, Kucuk O, et al. Genistein potentiates inhibition of tumor growth by radiation in a prostate cancer orthotopic model. *Mol Cancer Ther* 2004;3:1271-1279.
36. Strickley RG. Solubilizing excipients in oral and injectable formulations. *Pharm Res* 2004;21:201-230.
37. Chaturvedi MM, Mukhopadhyay A, Aggarwal BB. Assay for redox-sensitive transcription factor. *Methods Enzymol* 2000;319:585-602.
38. Aggarwal BB, Ichikawa H. Molecular targets and anticancer potential of indole-3-carbinol and its derivatives. *Cell Cycle* 2005;4:1201-1215.
39. Brandi G, Paiardini M, Cervasi B, et al. A new indole-3-carbinol tetrameric derivative inhibits cyclin-dependent kinase 6 expression, and induces G1 cell cycle arrest in both estrogen-dependent and estrogen-independent breast cancer cell lines. *Cancer Res* 2003;63:4028-4036.

40. Riby JE, Chang GH, Firestone GL, et al. Ligand-independent activation of estrogen receptor function by 3, 3'-diindolylmethane in human breast cancer cells. *Biochem Pharmacol* 2000;60:167-177.
41. Riby JE, Feng C, Chang YC, et al. The major cyclic trimeric product of indole-3-carbinol is a strong agonist of the estrogen receptor signaling pathway. *Biochemistry* 2000;39:910-918.
42. Shertzer HG. Indole-3-carbinol protects against covalent binding of benzo[a]pyrene and N-nitrosodimethylamine metabolites to mouse liver macromolecules. *Chem Biol Interact* 1984;48:81-90.
43. Shertzer HG, Senft AP. The micronutrient indole-3-carbinol: implications for disease and chemoprevention. *Drug Metabol Drug Interact* 2000;17:159-188.
44. Firestone GL, Bjeldanes LF. Indole-3-carbinol and 3-3'-diindolylmethane antiproliferative signaling pathways control cell-cycle gene transcription in human breast cancer cells by regulating promoter-Sp1 transcription factor interactions. *J Nutr* 2003;133:2448S-2455S.
45. Meng Q, Goldberg ID, Rosen EM, et al. Inhibitory effects of Indole-3-carbinol on invasion and migration in human breast cancer cells. *Breast Cancer Res Treat* 2000;63:147-152.
46. Wu HT, Lin SH, Chen YH. Inhibition of cell proliferation and in vitro markers of angiogenesis by indole-3-carbinol, a major indole metabolite present in cruciferous vegetables. *J Agric Food Chem* 2005;53:5164-5169.
47. Nemeth JA, Yousif R, Herzog M, et al. Matrix metalloproteinase activity, bone matrix turnover, and tumor cell proliferation in prostate cancer bone metastasis. *J Natl Cancer Inst* 2002;94:17-25.
48. Hernandez-Lopez C, Varas A, Sacedon R, et al. Stromal cell-derived factor 1/CXCR4 signaling is critical for early human T-cell development. *Blood* 2002;99:546-554.
49. Curran S, Murray GI. Matrix metalloproteinases: molecular aspects of their roles in tumour invasion and metastasis. *Eur J Cancer* 2000;36:1621-1630.
50. Dong Z, Bonfil RD, Chinni S, et al. Matrix metalloproteinase activity and osteoclasts in experimental prostate cancer bone metastasis tissue. *Am J Pathol* 2005;166:1173-1186.
51. John A, Tuszynski G. The role of matrix metalloproteinases in tumor angiogenesis and tumor metastasis. *Pathol Oncol Res* 2001;7:14-23.
52. Li Y, Che M, Bhagat S, et al. Regulation of Gene Expression and Inhibition of Experimental Prostate Cancer Bone Metastasis by Dietary Genistein. *Neoplasia* 2004;6:354-363.
53. Li Y, Kucuk O, Hussain M, et al. Antitumor and antimetastatic activities of docetaxel are enhanced by genistein through regulation of osteoprotegerin/receptor activator of nuclear

- factor-kappaB (RANK)/RANK ligand/MMP-9 signaling in prostate cancer. *Cancer Res* 2006;66:4816-4825.
54. McCawley LJ, Matrisian LM. Matrix metalloproteinases: they're not just for matrix anymore! *Curr Opin Cell Biol* 2001;13:534-540.
55. Przybylowska K, Kluczna A, Zadrozny M, et al. Polymorphisms of the promoter regions of matrix metalloproteinases genes MMP-1 and MMP-9 in breast cancer. *Breast Cancer Res Treat* 2006;95:65-72.
56. Sato H, Takino T, Okada Y, et al. A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* 1994;370:61-65.
57. Waas ET, Lomme RM, DeGroot J, et al. Tissue levels of active matrix metalloproteinase-2 and -9 in colorectal cancer. *Br J Cancer* 2002;86:1876-1883.
58. Wang M, Wang T, Liu S, et al. The expression of matrix metalloproteinase-2 and -9 in human gliomas of different pathological grades. *Brain Tumor Pathol* 2003;20:65-72.
59. Goldberg-Bittman L, Neumark E, Sagi-Assif O, et al. The expression of the chemokine receptor CXCR3 and its ligand, CXCL10, in human breast adenocarcinoma cell lines. *Immunol Lett* 2004;92:171-178.
60. Smith MC, Luker KE, Garbow JR, et al. CXCR4 regulates growth of both primary and metastatic breast cancer. *Cancer Res* 2004;64:8604-8612.
61. Cover CM, Hsieh SJ, Tran SH, et al. Indole-3-carbinol inhibits the expression of cyclin-dependent kinase-6 and induces a G1 cell cycle arrest of human breast cancer cells independent of estrogen receptor signaling. *J Biol Chem* 1998;273:3838-3847.
62. Garcia HH, Brar GA, Nguyen DH, et al. Indole-3-carbinol (I3C) inhibits cyclin-dependent kinase-2 function in human breast cancer cells by regulating the size distribution, associated cyclin E forms, and subcellular localization of the CDK2 protein complex. *J Biol Chem* 2005;280:8756-8764.
63. Herraiz T, Galisteo J. Endogenous and dietary indoles: a class of antioxidants and radical scavengers in the ABTS assay. *Free Radic Res* 2004;38:323-331.
64. Manson MM, Farmer PB, Gescher A, et al. Innovative agents in cancer prevention. *Recent Results Cancer Res* 2005;166:257-275.
65. Liang Z, Wu T, Lou H, et al. Inhibition of breast cancer metastasis by selective synthetic polypeptide against CXCR4. *Cancer Res* 2004;64:4302-4308.
66. Allinen M, Beroukhi R, Cai L, et al. Molecular characterization of the tumor microenvironment in breast cancer. *Cancer Cell* 2004;6:17-32.

67. Ricca A, Biroccio A, Del Bufalo D, et al. bcl-2 over-expression enhances NF-kappaB activity and induces mmp-9 transcription in human MCF7(ADR) breast-cancer cells. *Int J Cancer* 2000;86:188-196.
68. Takada Y, Andreeff M, Aggarwal BB. Indole-3-carbinol suppresses NF-kappaB and IkappaBalpha kinase activation, causing inhibition of expression of NF-kappaB-regulated antiapoptotic and metastatic gene products and enhancement of apoptosis in myeloid and leukemia cells. *Blood* 2005;106:641-649.

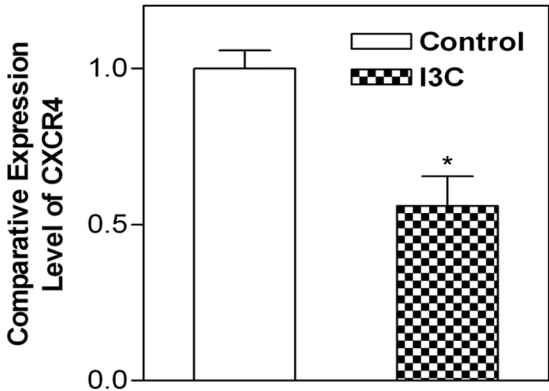


Fig 1

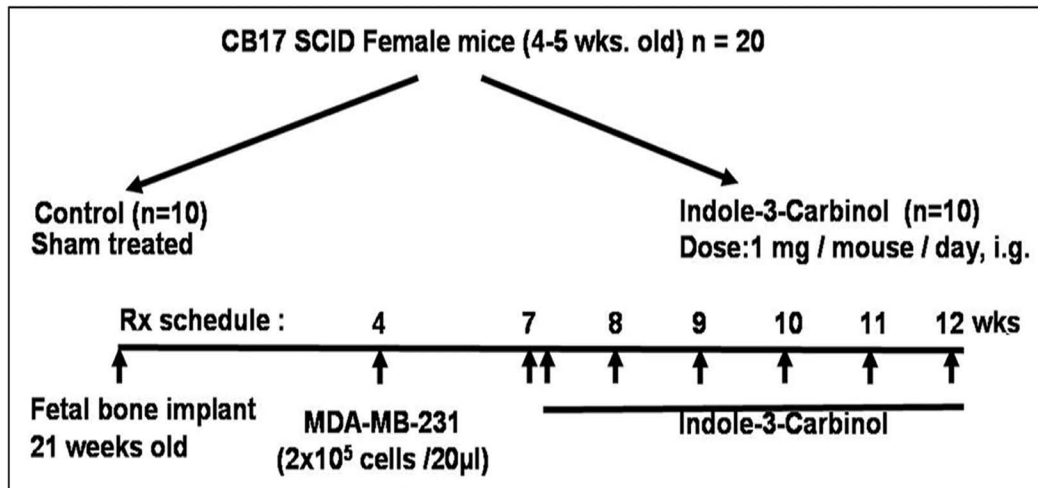


Fig 2

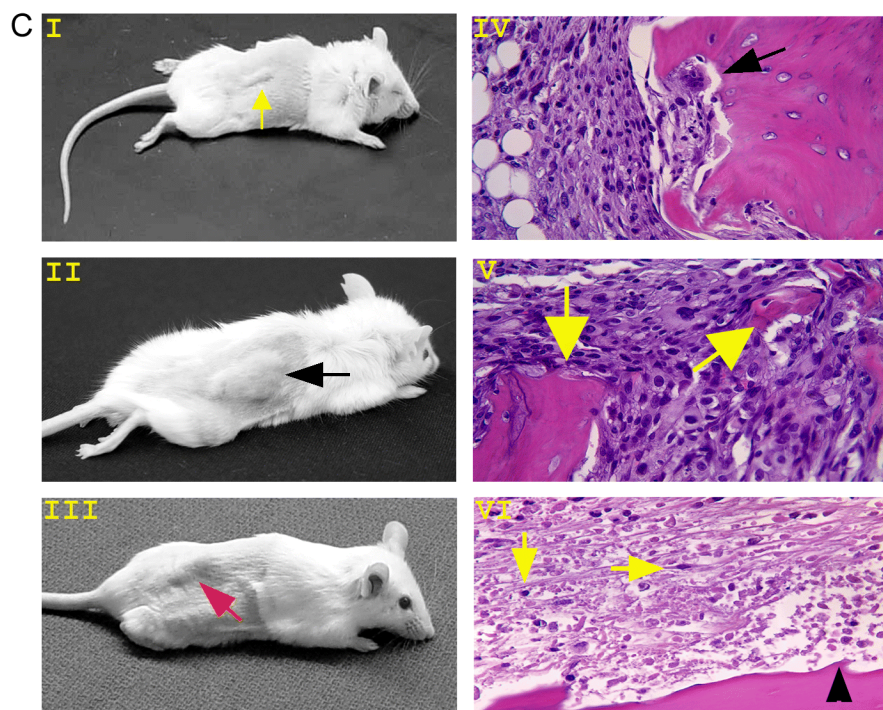
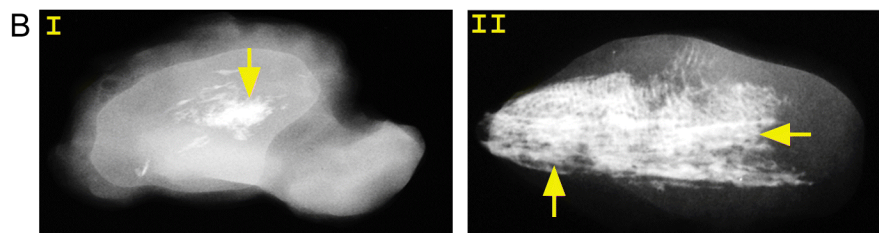
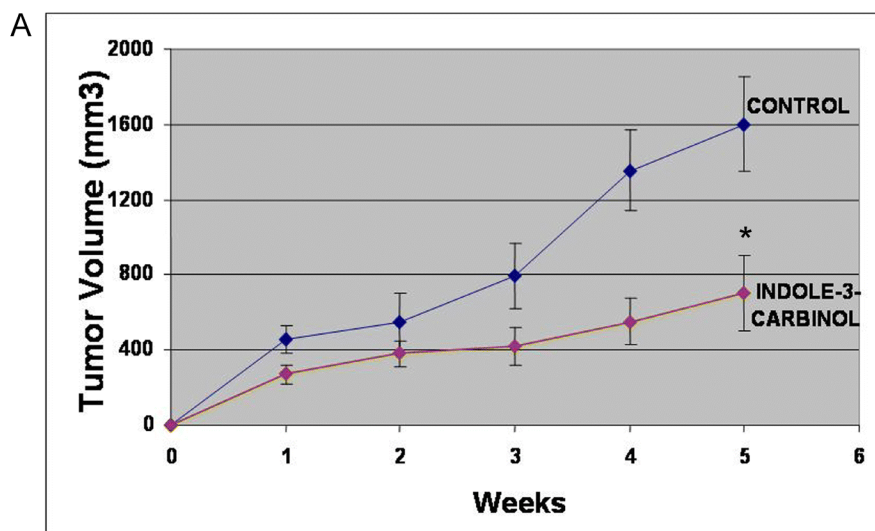


Fig 3

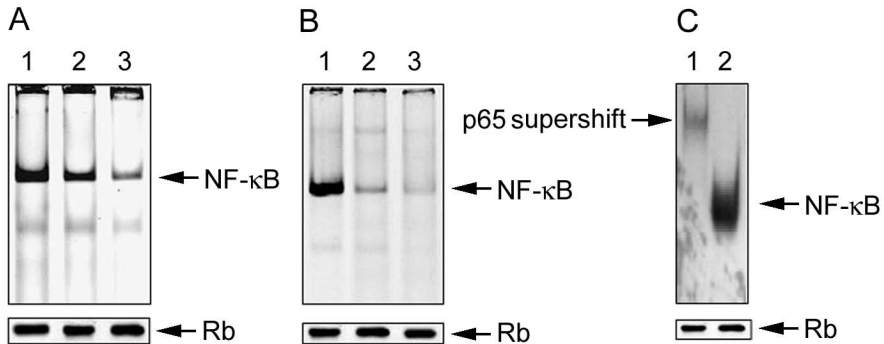
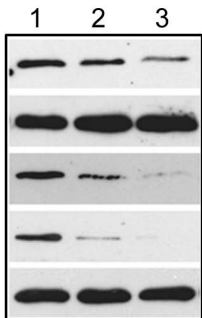


Fig 4

A



B

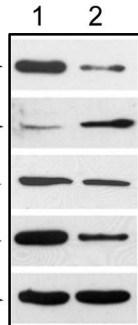


Fig 5

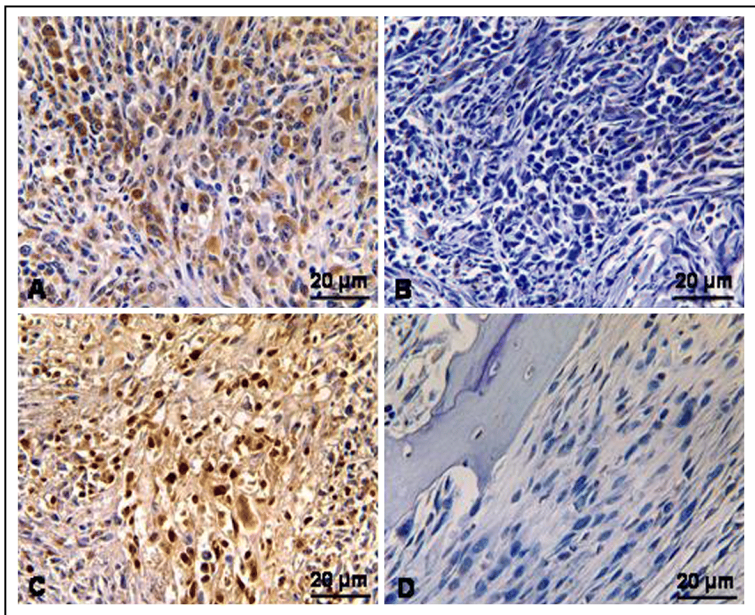


Fig 6

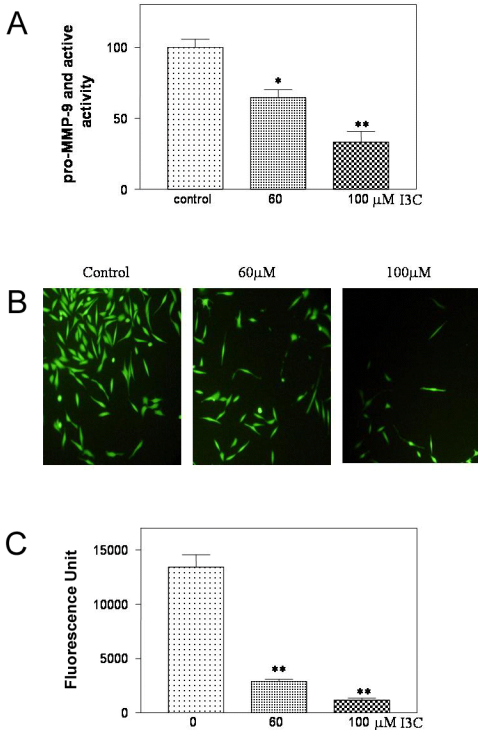


Fig 7

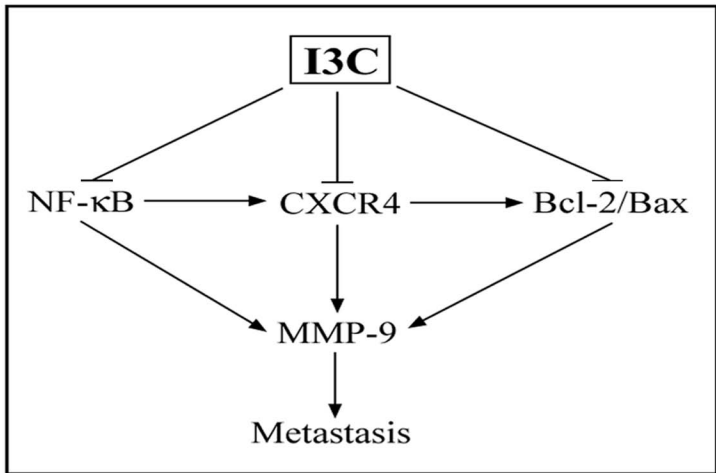


Fig 8